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[54] ISOLATED GENE ENCODING AN ENZYME WITH UDP-GLUCOSE PYROPHOSPHORYLASE AND PHOSPHOGLUCOMUTASE ACTIVITIES FROM CYCLOTELLA CRYPTICA

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[52] U.S. Cl. 435/257.2; 536/23.6; 536/23.2; 435/6; 435/69.1; 435/71.1; 435/194; 435/252.3; 435/257.1; 435/320.1

[58] Field of Search 536/23.2, 23.6; 435/6, 69.1, 71.1, 194, 252.3, 257.1, 257.2, 320.1

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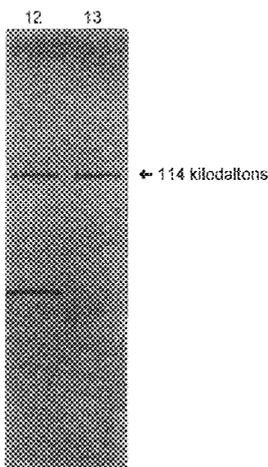
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[57] ABSTRACT

The present invention relates to a cloned gene which encodes an enzyme, the purified enzyme, and the applications and products resulting from the use of the gene and enzyme. The gene, isolated from Cyclotella cryptica, encodes a multifunctional enzyme that has both UDP-glucose pyrophosphorylase and phosphoglucomutase activities.

11 Claims, 4 Drawing Sheets



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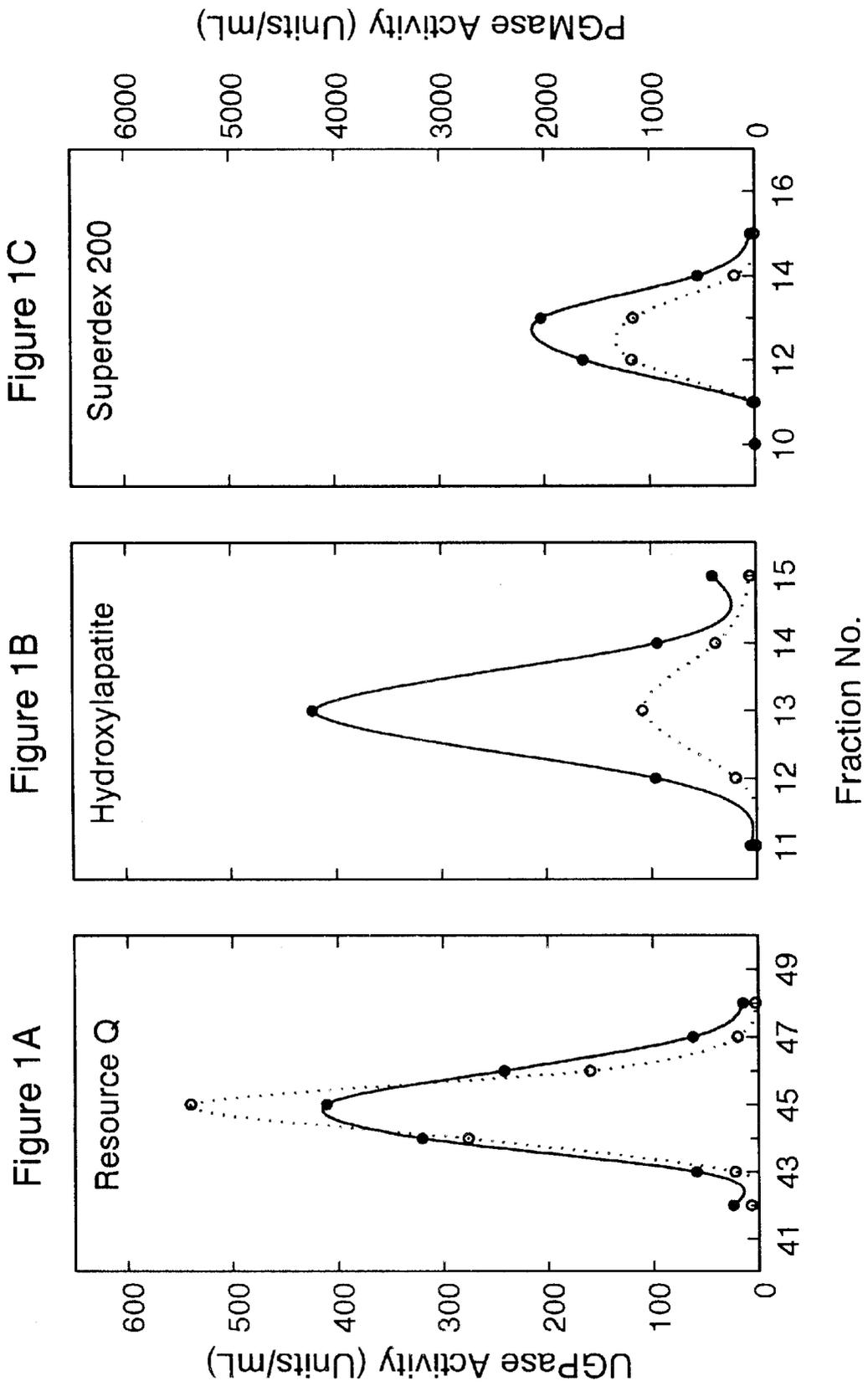


Figure 2

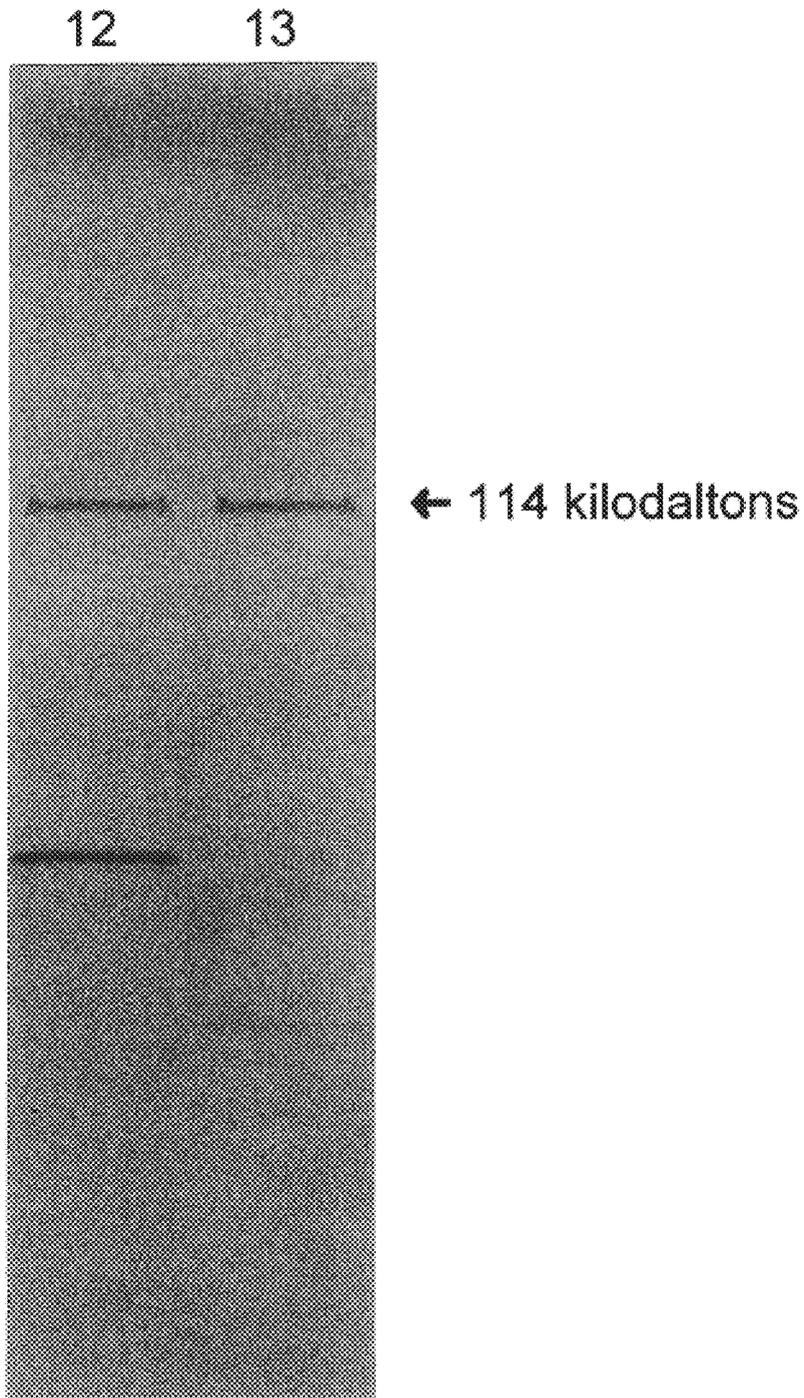


Figure 3A

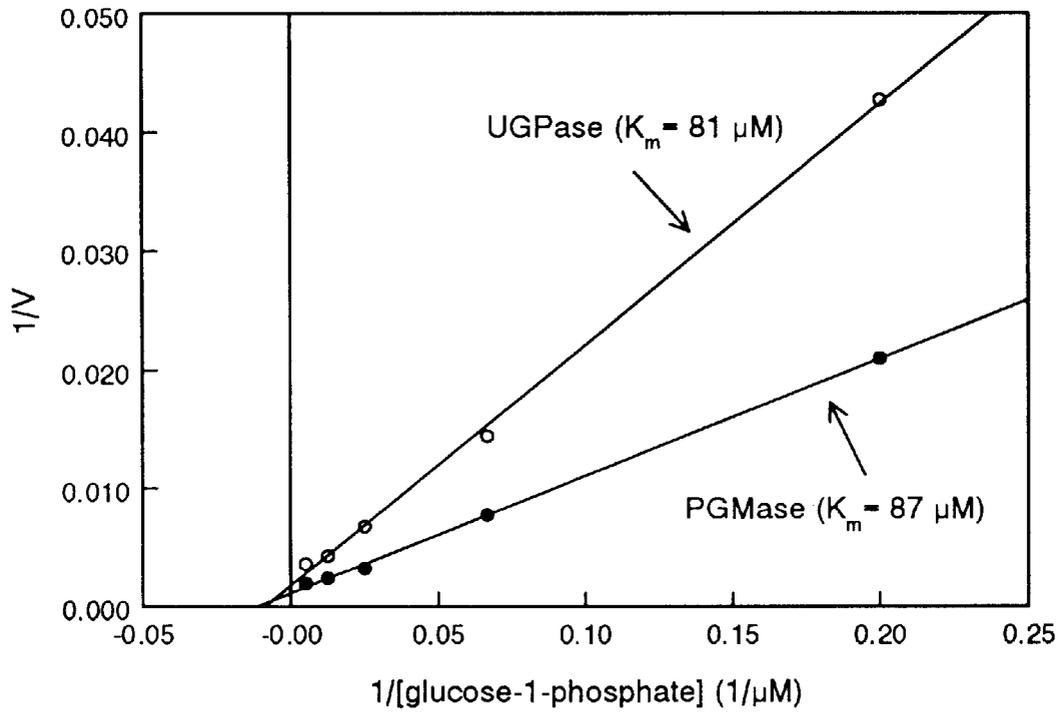


Figure 3B

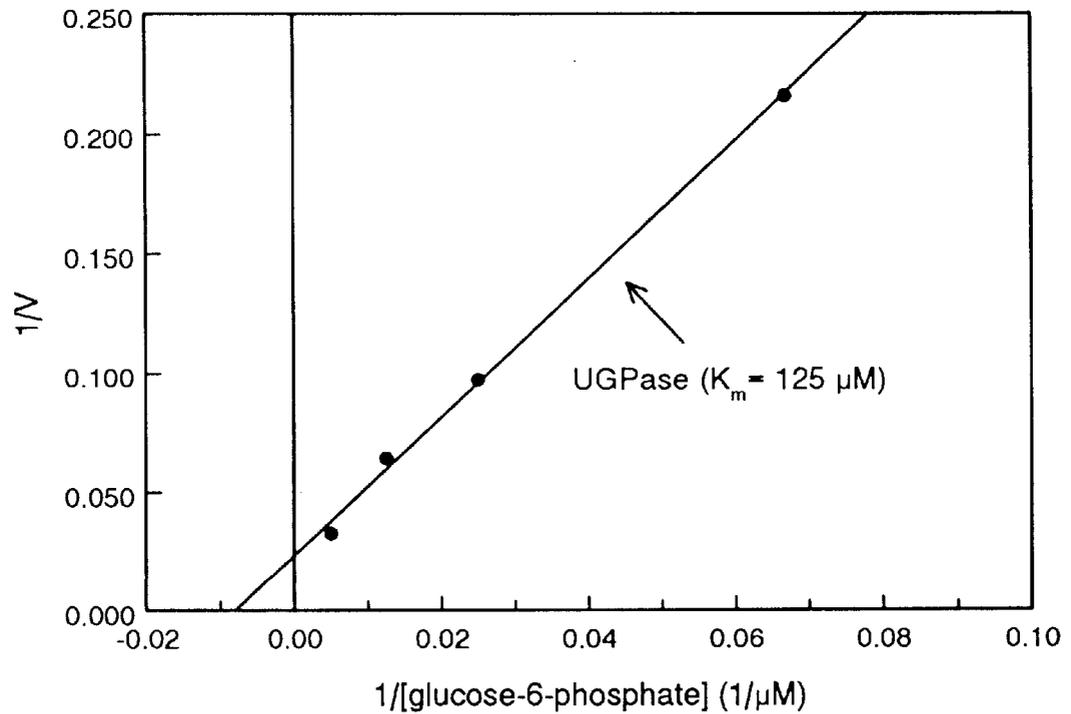


Figure 4B

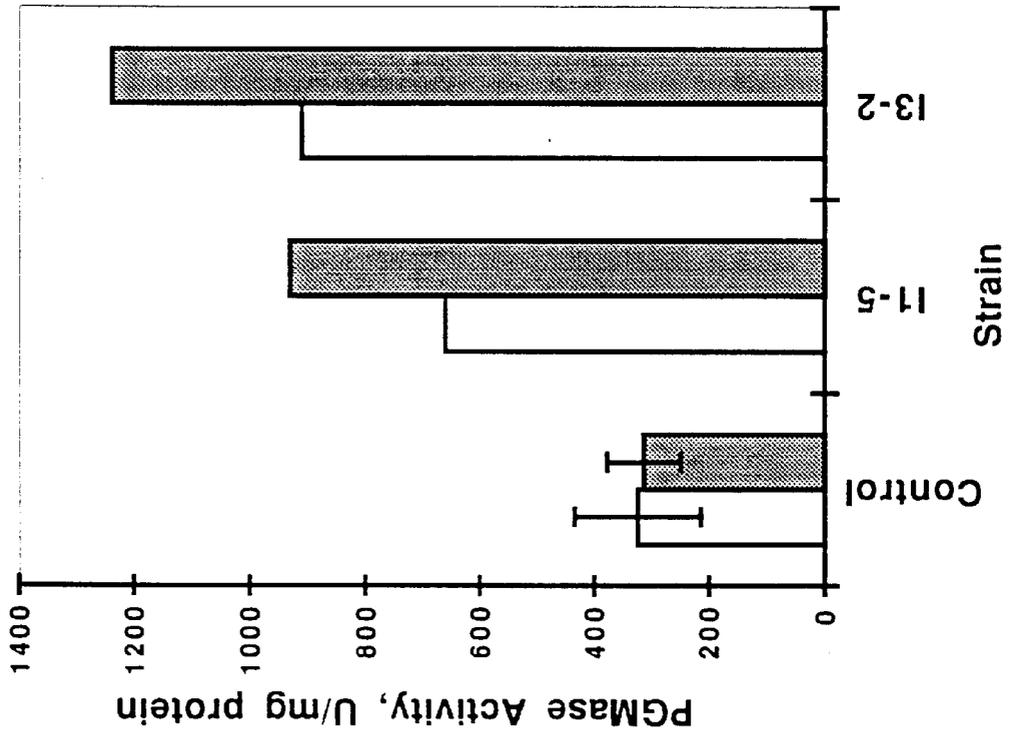
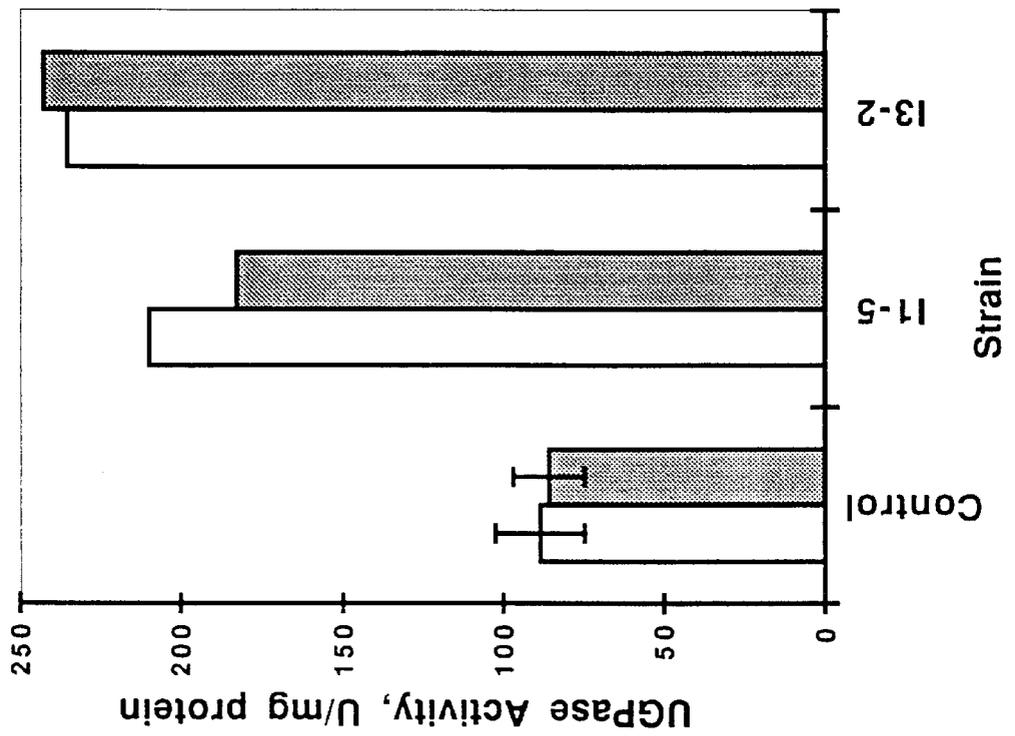


Figure 4A



**ISOLATED GENE ENCODING AN ENZYME
WITH UDP-GLUCOSE
PYROPHOSPHORYLASE AND
PHOSPHOGLUCOMUTASE ACTIVITIES
FROM *CYCLOTELLA CRYPTICA***

The United States Government has rights in this invention under Contract No. DE-AC36-83CH10093 between the United States Department of Energy and the National Renewable Energy Laboratory, a Division of the Midwest Research Institute.

FIELD OF THE INVENTION

The invention relates to a cloned gene encoding an enzyme involved in the metabolism of carbohydrates in algae.

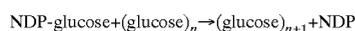
BACKGROUND OF THE INVENTION

The present invention relates to a cloned gene which encodes an enzyme, the purified enzyme, and the applications and products resulting from the use of the gene and enzyme.

Polysaccharides, sugars, and lipids are the primary constituents of many food and industrial products. These products are central to human and animal nutrition and therefore have significant commercial value. Crop plants are a primary source of these compounds. Numerous crop improvement efforts around the world are focused on altering the amounts and ratios of these compounds in various species.

Carbohydrates are a major form of energy storage in plants and animals. Long chain storage carbohydrates can take many forms, but most commonly are polymers of glucose molecules; these polymers are referred to as glucans. The glucose molecules in these polysaccharides can be joined together through a variety of different linkages. The storage polysaccharides starch and glycogen are α -1,4, linked glucans. Starch is the primary storage glucan in higher plants, and starch reserves in seeds comprise a major source of carbohydrate for human and animal nutrition and for the production of certain commodity chemicals such as ethanol. Glycogen is a common energy storage compound for animals and many microbes. β -1,3 linked glucans are present as storage carbohydrates in numerous algal taxa, including laminarin (Phaeophyceae), chrysolaminarin (Bacillariophyceae and Chrysophyceae), and paramylon (Euglenophyceae). The β -1,4 linked glucans include the structural polymer cellulose, which is one of the most abundant biological polymers on earth.

The biosynthetic pathways of all of these glucans share a common mechanistic theme: the basic building block (i.e., the substrate for chain elongation) is an activated glucose molecule. Activation is achieved by conjugation of glucose to a nucleoside diphosphate (NDP) such as uridine diphosphate (UDP) or adenosine diphosphate (ADP). Actual synthesis of the polymer is mediated by synthase enzymes (e.g., starch synthase), which utilize NDP-glucose molecules as substrates. The general reaction catalyzed by glucan synthases is shown below:



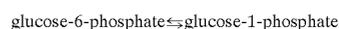
Sucrose is the principal carbon transport molecule in higher plants and is an important food ingredient. The biosynthesis of sucrose also utilizes a nucleoside diphosphate-activated glucose molecule, generally UDP-glucose. The biosynthesis of many other disaccharides and complex sugars also uses NDP-glucose as a substrate.

Specific enzymes are responsible for formation of the NDP-glucose molecules that are used for glucan and complex sugar biosynthesis. UDP-glucose is formed through the action of the enzyme UDP-glucose pyrophosphorylase (E.C. 2.7.7.9; also known as glucose-1-phosphate uridylyltransferase, and hereinafter referred to as UGPase). The reaction catalyzed by UGPase is as follows:



where UTP is uridine triphosphate and PP_i is pyrophosphate. The subsequent hydrolysis of pyrophosphate to orthophosphate is responsible for driving the reaction toward the formation of UDP-glucose. UGPase genes have been cloned from disparate sources, including potato (Katsube et al., J. Biochem. 108: 321-326 (1990)), human (Peng and Chang, FEBS Lett. 329:153-158 (1993)), and yeast (Purnelle et al., Yeast 8:977-986 (1992)). The UGPase enzyme has also been purified from various sources, including barley (Elling and Kula, J. Biotechnol. 34:157-163 (1994)), rice (Kimura et al., Plant Physiol. Biochem. 30:683-693 (1992)), and potato (Nakano et al., J. Biochem. 106:528-532 (1989)).

The substrate for UGPase, glucose-1-phosphate, is synthesized from glucose-6-phosphate through the action of the enzyme phosphoglucomutase (E.C. 5.4.2.2; hereinafter referred to as PGMase), as shown below:



The PGMase enzyme has been purified from many sources, including pea (Galloway and Dugger, Physiol. Plant. 92:479-486 (1994)), human (Fazi et al., Prep. Biochem. 20:219-240 (1990)), and *Lactobacillus* (Marechal et al., Arch. Biochem. Biophys. 228:592-599 (1984)). The PGMase gene has also been cloned from a number of sources, including human (Putt et al., Biochem. J. 296:417-422 (1993)), yeast (GenBank accession no. X72016), and *E. coli* (GenBank accession no. U08369).

The present invention concerns a novel carbohydrate biosynthesis gene isolated from a microalga. Microalgae are defined as unicellular, eukaryotic algae. Although their current biotechnological utilization is primarily for the production of high value specialty products, microalgae have very high productivity rates that could support the large-scale, commercial production of lower value carbohydrates and lipids. One of the species under consideration for such uses is the centric diatom *Cyclotella cryptica*. This organism grows naturally in salt water and has been shown to be highly productive in outdoor culture (Weissman and Tillett, NREL/TP-232-4147:32-56, (1989)). *C. cryptica* is under consideration for the production of alternative fuels because lipids can comprise up to 40-60% of the cellular dry weight when cells are grown under nutrient-limiting conditions. These lipids are similar in composition to the triacylglycerols produced by oilseed crops and can be readily converted, via transesterification with a simple alcohol, into a diesel fuel replacement.

In addition to its ability to accumulate lipids, *C. cryptica* produces a substantial amount of carbohydrate. Approximately 20-30% of the dry weight of *C. cryptica* cells consists of a β -1,3 linked glucan referred to as chrysolaminarin (Roessler, J. Phycol. 23:494-498 (1987)). This glucan accumulates in all growth phases and decreases only slightly upon the induction of lipid accumulation in nutrient-deficient cells. Thus, this carbohydrate constitutes a significant sink for fixed carbon, and therefore competes for carbon substrates with the lipid biosynthetic pathway. Roessler (Roessler, J. Phycol. 23:494-498 (1987)) demonstrated pre-

viously that the precursor for chrysolaminarin biosynthesis in *C. cryptica* is UDP-glucose, and that UGPase enzyme activity was present in extracts of *C. cryptica* cells. The UDP-glucose produced by UGPase is a substrate for the enzyme chrysolaminarin synthase, which adds glucose units successively onto the growing carbohydrate polymer. In contrast, PGMase in *C. cryptica* has not been characterized.

The instant invention is directed to the isolation of a gene from *C. cryptica* that encodes a multifunctional enzyme that has both UGPase and PGMase activities. This is the first report of the isolation of a gene encoding either of these enzymes from an alga. The fact that UGPase and PGMase domains are both present on a single polypeptide chain could not have been anticipated; these activities have never before been reported to exist together on a single protein. Uttaro and Ugalde (Uttaro and Ugalde, *Gene* 150:117-122 (1994)) reported a chromosomal cluster in the bacterium *Agrobacterium tumefaciens* that encodes ADP-glucose pyrophosphorylase, glycogen synthase, and PGMase; however, the three activities are encoded by three separate open reading frames, and are consequently found on three separate proteins. The presence of UGPase and PGMase on a single polypeptide chain could have significant advantages both in terms of more favorable reaction kinetics and because a single gene can be inserted into an organism via genetic engineering to affect two enzymatic functions simultaneously. Artificial polypeptide fusions have been shown to have kinetic advantages in other systems (for example, Tamada et al., *Bioconjugate Chem.* 5:660-665 (1994)). A naturally-occurring fusion may exhibit even greater kinetic advantages than man-made fusions, in that evolutionary selective pressure can result in functionally superior enzymes.

SUMMARY OF THE INVENTION

An object of this invention is to affect the production of storage compounds in biological organisms.

Another object of this invention is to develop plant, algal, or microbial species that produce more or less carbohydrate, sugar (including sucrose), or lipid.

Another object of this invention is to provide a protein having two enzymatic activities.

Another object of this invention is to provide a protein having two enzymatic activities wherein the activities are UGPase and PGMase.

Another object of this invention is to provide a single gene encoding a protein having two enzymatic activities.

Another object of this invention is to provide a single gene encoding a protein having two enzymatic activities wherein the activities are UGPase and PGMase.

Another object of this invention is to develop strains of *C. cryptica* and related microalgae that produce more lipid through the inactivation or inhibition of competing carbohydrate biosynthetic pathways.

To accomplish these goals, a gene encoding UGPase and PGMase has been isolated and cloned from *C. cryptica*. In this organism, the two enzymes are present as separate domains on a single polypeptide chain, which is encoded by a single gene. The protein encoded by this gene has been purified to near homogeneity and has been shown to carry out both enzymatic activities *in vitro*. The gene could be overexpressed in plants, algae, or other microorganisms in order to increase PGMase and UGPase activities *in vivo*, thus potentially altering the amounts or ratios of carbohydrate, sugar (including sucrose), or lipid produced. The expression of the gene could be inhibited in *C. cryptica*

or a related species by introduction of antisense, ribozyme, co-suppression, or other constructs that are based on the DNA sequence of this gene.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A illustrates UGPase and PGMase activity in 1.5 mL fractions eluting from a 6 mL Resource Q ion exchange chromatography column (Pharmacia Biotech, Inc.; Piscataway, N.J.). UGPase activity is represented by a solid line, and PGMase activity is represented by a dotted line. One Unit of activity is defined as one nmol of product formed per minute. Note that the activities of UGPase and PGMase co-elute.

FIG. 1B illustrates UGPase and PGMase activity in one mL fractions eluting from a 7x52 mm Bio-Scale CHT2-1 hydroxylapatite chromatography column (BioRad Laboratories; Hercules, Calif.). UGPase activity is represented by a solid line, and PGMase activity is represented by a dotted line. One Unit of activity is defined as one nmol of product formed per minute. Note that the activities of UGPase and PGMase co-elute.

FIG. 1C illustrates UGPase and PGMase activity in one mL fractions eluting from a 1x30 cm Superdex 200 gel filtration chromatography column (Pharmacia Biotech, Inc.; Piscataway, N.J.). UGPase activity is represented by a solid line, and PGMase activity is represented by a dotted line. One Unit of activity is defined as one nmol of product formed per minute. Note that the activities of UGPase and PGMase co-elute.

FIG. 2 is a photograph of a sodium dodecyl sulfate-polyacrylamide gel (10% polyacrylamide, w/w) that has been stained with Coomassie Blue R-250, showing a substantially purified preparation of the UGPase/PGMase protein. The molecular weight of the purified protein was estimated to be 114 kilodaltons based on the migration distances of protein standards. The number above each lane represents the fraction number from the gel filtration chromatography step illustrated in FIG. 1C above.

FIG. 3A illustrates double reciprocal plots of UGPase and PGMase activity versus glucose-1-phosphate concentration. Velocity (V) is expressed in Units/mL. UGPase activity is represented by open circles, and PGMase activity is represented by filled circles.

FIG. 3B illustrates a double reciprocal plot of UGPase activity versus glucose-6-phosphate concentration. Velocity (V) is expressed in Units/mL.

FIG. 4A illustrates overexpression of the UGPase/PGMase (*upp1*) gene in recombinant *C. cryptica* cells, as determined via the measurement of UGPase activity. The white and gray bars represent the results from two independent experiments. Error bars for the control data indicate \pm one standard deviation around the mean for six independent strains transformed with a control plasmid. One Unit (U) of activity is defined as one nmol of product formed per minute.

FIG. 4B illustrates overexpression of the *upp1* gene in recombinant *C. cryptica* cells, as determined via the measurement of PGMase activity. The white and gray bars represent the results from two independent experiments. Error bars for the control data indicate \pm one standard deviation around the mean for six independent strains transformed with a control plasmid. One Unit (U) of activity is defined as one nmol of product formed per minute.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The gene for UGPase and PGMase (hereinafter referred to as the *upp1* gene) from *C. cryptica* encodes a polypeptide

that is 1056 amino acids in length and that has a predicted molecular weight of 114.4 kilodaltons. UGPase and PGMase enzyme functions are both present on this single polypeptide. The genomic sequence contains three introns, which are 290, 103, and 76 base pairs (bp) in length.

Standard cloning techniques were performed as described in Sambrook et al. (*Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press (1989)), and the terminology herein is used as defined in that reference. Unless specifically defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described.

EXAMPLE

For the experiments below, the strain *Cyclotella cryptica* T13L was used. This strain was obtained from the Bigelow Laboratory Culture Collection of Marine Phytoplankton, West Boothbay Harbor, Me. *C. cryptica* was cultured as described in Roessler (Roessler, J. Phycol. 24: 394-400 (1988)).

Cloning of the upp1 Gene

The *C. cryptica* upp1 gene was isolated as follows. A fragment of the gene was first produced by the polymerase chain reaction (PCR) using degenerate oligonucleotide primers that were based on conserved sequences from other known UGPase genes, specifically potato (Katsube et al., J. Biochem. 108:321-326 (1990)), human (Peng and Chang, FEBS Lett. 329:153-158 (1993)), yeast (Purnelle et al., Yeast 8: 977 986 (1992)), and Dictyostelium (Ragheb and Dottin, Nucleic Acids Res. 15:3891-3906 (1987)). Three primers were designed that are referred to as UP2, UP3, and UP4. UP3 is an upstream primer; UP2 and UP4 are downstream primers, with UP4 being further downstream than UP2. The DNA sequences for the primers are given as follows (using IUPAC codes):

UP2: 5'-ARRTTRTINGTRITRAA (17-mer, 128-fold degenerate) also identified herein as SEQ ID NO:1.

UP3: 5'-TGGTAYCCNCCNGGWC (17-mer, 64-fold degenerate) also identified herein as SEQ ID NO:2.

UP4: 5'-GCNGTYTCNARYTG (14-mer, 128-fold degenerate) also identified herein as SEQ ID NO:3.

A nested PCR strategy was used to isolate a portion of the upp1 gene. First, primers UP3 and UP4 were used in a PCR amplification with the following conditions; a 20 μ l PCR reaction contained 36 ng of total DNA from *C. cryptica*, 2.5 μ M of each primer, 10 mM Tris-Cl (pH 8.3), 50 KCl, 1.5 mM MgCl₂, 0.2 mM dNTPs, and 0.75 units of Taq DNA polymerase (Perkin Elmer-Cetus; Norwalk, Conn.). Total DNA was isolated from *C. cryptica* by the method of Jarvis et al. (Jarvis et al., J. Phycol. 28:356-362 (1992)). The following thermal cycle was used; Step 1, 94° C. for 2 min; Step 2, 45° C. for 30 sec; Step 3, 72° C. for 1 min; Step 4, 94° C. for 30 sec; Step 5, 45° C. for 30 sec; Step 6, repeat steps 3 to 5 for 30 times total; and Step 7, 72° C. for 10 min. The products of this reaction were analyzed on a 2.8% agarose gel, which indicated a large number of non-specific DNA products.

The products of this first amplification were subjected to a secondary, nested amplification using primers UP3 and UP2. The conditions for this amplification were identical to the first except that 0.1 μ l of the first reaction was used as the template, and only 20 cycles were run instead of 30. The

products of this reaction were also analyzed by gel electrophoresis and exhibited 10 to 15 distinct DNA products. One of the DNA products was 338 bp in length, which was the expected size based on the sequences of the other known UGPase genes. This fragment was cut from the gel, purified by use of a "Gene Clean" kit (BIO 101; La Jolla, Calif.), and reamplified with primers UP2 and UP3 using the previous reamplification conditions.

The product of this third reaction was subcloned into the plasmid pCRII (Invitrogen; San Diego, Calif.) according to the manufacturer's instructions, and the resulting product was used to transform *E. coli* INV α F'. The cloned fragment was sequenced by the double-stranded sequencing protocol of Kraft et al. (Kraft et al., Biotechniques 6:544-546 (1988)). The 338-bp DNA fragment contained 304 bp of amplified *C. cryptica*-derived sequence; 34 bp were derived from the amplification primers. This *C. cryptica* DNA sequence is identified herein as SEQ ID NO:4. The deduced amino acid sequence of this fragment (identified herein as SEQ ID NO:5) exhibited 37% identity with the corresponding sequence of potato UGPase, thereby confirming that a *C. cryptica* UGPase gene fragment had been purified.

The cloned PCR product was subsequently used as a probe to isolate a lambda clone containing the entire *C. cryptica* UGPase gene. The lambda library used was constructed as described in Roessler and Ohlrogge (Roessler and Ohlrogge, J. Biol. Chem. 268:19254-19259 (1993)). Filter lifts of the library were screened with the cloned 338-bp UGPase gene fragment that was labeled via PCR with digoxigenin using the "Genius" non-radioactive labeling and detection system (Boehringer Mannheim Corporation; Indianapolis, Ind.) according to the manufacturer's instructions. Several positive clones were isolated in this manner. Restriction mapping and subcloning were used to identify a DNA segment shared by all of the positive clones. Sequencing of this DNA segment revealed that it contained a single long open reading frame, the 5' end of which showed homology to known UGPase genes and the 3' end of which showed homology to known PGMase genes. PGMase and UGPase are thought to catalyze successive steps in the chrysolaminarin biosynthesis pathway. The isolation of a naturally occurring fusion of these two genes is, to our knowledge, unprecedented, and this result could not have been anticipated.

Three introns were identified within the upp1 coding region. Exact splice junctions were determined by sequencing the PCR products made from *C. cryptica* cDNA. RNA was isolated as described in Roessler and Ohlrogge (Roessler and Ohlrogge, J. Biol. Chem. 268:19254-19259 (1993)), and subjected to reverse transcription and PCR using a kit (GeneAmp RNA PCR Kit; Perkin-Elmer Cetus; Norwalk, Conn.). Gene-specific primers based on the *C. cryptica* upp1 gene sequence were used. The DNA products of these amplifications were subcloned into pCRII and sequenced as described above. The three introns identified by this method are 290, 103, and 76 bp in length.

The DNA sequence of the upp1 gene, from the start codon through the stop codon, is identified herein as SEQ ID NO:6. The positions of the three introns are noted. The genomic sequence including the start and stop codons is 3640 bp in length; removal of the introns yields a 3171-bp coding sequence. The deduced amino acid sequence of the UGPase/PGMase protein is identified herein as SEQ ID NO:7. The predicted polypeptide is 1056 amino acids in length with a molecular weight of 114.4 kilodaltons. Based on similarities to known sequences, the boundary between the UGPase and PGMase domains is located approximately at position 480 in the amino acid sequence.

Purification of the UGPase/PGMase protein

To further characterize the protein encoded by the cloned *upp1* gene, several sequential column chromatographic procedures were carried out to purify the native UGPase/PGMase protein. The cells in one liter of an exponential phase culture were harvested by centrifugation at 15,000×g for 10 minutes, followed by washing with 8 mL of Buffer A (50 mM HEPES buffer, pH 7.5, containing 2 mM dithiothreitol) and repelleting by another centrifugation step. The cells were resuspended in 8 mL of Buffer A, frozen in liquid N₂, and then stored at -80° C. Cell-free extracts were prepared by subjecting the cells to three rounds of freezing in liquid N₂ and thawing at 30° C., followed by centrifugation at 37,000×g for 20 minutes. The extract was then filtered through a 0.2 μm filter.

Ion exchange chromatography was carried out by loading 4 mL of the filtered extract onto a 6 mL Resource Q anion exchange column (Pharmacia Biotech, Inc.; Piscataway, N.J.), followed by elution with a 120 mL linear gradient from 0 to 0.5M NaCl in Buffer A. Fractions (1.5 mL) were collected and independently assayed for the presence of UGPase and PGMase activities as described below. As shown in FIG. 1A, UGPase and PGMase activities were found in the same fractions; the peak activities eluted at a NaCl concentration of approximately 270 to 300 mM.

The active fractions from the ion exchange chromatography step were combined and desalted by passage through a 20 mL Hi-Trap desalting column (Pharmacia Biotech, Inc.; Piscataway, N.J.) that had been equilibrated with Buffer A. The desalted solution was loaded onto a 7×52 mm Bio-Scale CHT2-1 hydroxylapatite column (BioRad Laboratories; Hercules, Calif.), and eluted with a 24 mL linear gradient from 0 to 0.5M sodium phosphate in Buffer A. One mL fractions were collected and independently assayed for the presence of UGPase and PGMase activities as described in the "Enzyme Assays" section below. As shown in FIG. 1B, UGPase and PGMase activities were found in the same fractions, which eluted at a sodium phosphate concentration of approximately 270 to 290 mM.

The active fractions from the hydroxylapatite chromatography step were combined and concentrated to 0.2 mL by the use of a Centricon-30 ultrafiltration device (Amicon, Inc.; Beverly, Mass.). This solution was loaded onto a 1×30 cm Superdex 200 gel filtration column (Pharmacia Biotech, Inc.; Piscataway, N.J.) that had been equilibrated with Buffer A. One mL fractions were collected and independently assayed for the presence of UGPase and PGMase activities. As shown in FIG. 1C, UGPase and PGMase activities were once again found in the same fractions.

The size exclusion chromatography fractions containing UGPase and PGMase activities were subjected to electrophoresis through an sodium dodecyl sulfate polyacrylamide gel (10% polyacrylamide, w/w) to determine the degree of purification. Thirty μL from each fraction were loaded per lane, and after electrophoresis the proteins in the gel were visualized by staining with Coomassie Blue R250. These results are shown in FIG. 2. Fractions number 12 and 13 had approximately equal activities; both of these fractions contained a 114 kilodalton protein that stained with equal intensity in both fractions. In fraction 13, this protein was purified to near homogeneity, providing strong evidence that this was the UGPase/PGMase protein encoded by the cloned *upp1* gene.

Enzyme Assays

UGPase activity was assayed by measuring the conversion of [¹⁴C]glucose-1-phosphate to [¹⁴C]UDP-glucose. The assay was carried out essentially as described by Roessler

(Roessler, J. Phycol. 24:394-400 (1988)), in which [¹⁴C] glucose-1-phosphate is incubated with UTP in the presence of UGPase for a specified period of time, after which the remaining substrate is dephosphorylated and the radioactively-labeled product ([¹⁴C]UDP-glucose) is bound onto DEAE-filter paper for liquid scintillation counting. The reaction mixture contained 50 mM Hepes buffer (pH 7.8), 5 mM MgCl₂, 1 mM UTP, and 0.5 mM [¹⁴C]glucose-1-phosphate (American Radiolabeled Chemicals, Inc.; St. Louis, Mo. at a specific activity of 1 mCi/mmol. Enzyme extract was added to tubes containing prewarmed reaction mixtures to begin the reaction, which was then allowed to proceed for 15 minutes at 30° C. The reaction was terminated by placing the tubes into boiling water for 3 minutes, followed by cooling on ice. For the dephosphorylation reaction, 0.5 units of bacterial alkaline phosphatase (Sigma Chemical Co.; St. Louis, Mo.; catalog no. P-4252) were added to each reaction and incubated for one hour at 30° C. The reaction mixtures were spotted onto separate 2.5 cm DE81 filter disks (Whatman Inc.; Fairfield, N.J.), which were then washed in five consecutive batches of water (200 mL each). The radioactivity of each filter was determined by liquid scintillation counting. Experiments utilizing glucose-6-phosphate for K_m determinations were performed in the same fashion except that [¹⁴C]glucose-6-phosphate (American Radiolabeled Chemicals, Inc.; St. Louis, Mo.) was used as the substrate and 10 μM glucose-1,6-diphosphate was included in the reactions.

PGMase activity was measured in a coupled assay in which glucose-6-phosphate is produced from glucose-1-phosphate via the action of PGMase, and then converted to 6-phosphogluconate through the action of exogenously added glucose-6-phosphate dehydrogenase with the subsequent reduction of NADP³⁰ to NADPH, which is measured spectrophotometrically by recording the change in absorbance at 340 nm. The reaction mixture contained 50 mM HEPES (pH 7.8), 5 mM MgCl₂, 1 mM NADP⁺, 0.5 mM glucose-1-phosphate, 10 μM glucose-1,6-diphosphate, one unit of glucose-6-phosphate dehydrogenase (Boehringer Mannheim Corporation; Indianapolis, Ind.; catalog no. 165875), and enzyme extract. The reaction was carried out at 25° C. for 2.5 minutes.

Amino Acid Sequencing of the Purified UGPase/PGMase Protein

The co-elution of UGPase and PGMase activities throughout the course of the protein purification procedure, along with the presence of a nearly pure 114 kilodalton polypeptide after the final size exclusion chromatography step, provided strong evidence that the protein product of the *upp1* gene had been purified. In order to provide absolute confirmation that the purified protein was the product of the cloned *upp1* gene, partial amino acid sequences were determined for peptides produced from the purified protein via proteolytic digestion. These sequences were then compared to the amino acid sequence deduced from the DNA sequence of the *upp1* gene.

The purified polypeptide that migrated at 114 kilodaltons during polyacrylamide gel electrophoresis was excised from the gel, and the gel slice was incubated in 0.1M ammonium bicarbonate containing 0.1% Tween 20 and lysyl peptidase. The resulting peptides were extracted twice with 200 μL of 50% acetonitrile containing 0.1% trifluoroacetic acid (TFA). The combined extracts were concentrated to 200 μL via vacuum centrifugation and diluted to 600 μL with 0.1% TFA. This sample was loaded onto a reverse phase high performance liquid chromatography column (2.1×30 mm Brownlee C18 Aquapore ODS column; Applied Biosystems,

Inc.; Foster City, Calif.) and separated with the following gradient: 5% B for 5 min, 5% B to 40% B over the next 45 min, and then 40% B to 70% B over the final 5 min; where A=0.1% TFA in water and B=0.09% TFA in acetonitrile. Two individual peptides resolved in this manner were sequenced via automated Edman degradation. The amino acid sequences determined for these two peptides are identified herein as SEQ ID NO:8 and SEQ ID NO:9. These sequences were identical to amino acid sequences predicted by the DNA sequence of the *upp1* gene, confirming that the substantially purified protein was the product of the *upp1* gene. SEQ ID NO:8 corresponds to positions 119 through 127 of the deduced amino acid sequence of the UGPase/PGMase protein (i.e., SEQ ID NO:7), whereas SEQ ID NO:9 corresponds to positions 711 through 729 of the deduced amino acid sequence of the UGPase/PGMase protein.

K Determinations

The K_m for glucose-1-phosphate was determined for both the UGPase and PGMase portions of the UGPase/PGMase enzyme via standard procedures (i.e., double reciprocal plots of enzyme activity versus glucose-1-phosphate concentration), using a highly purified preparation of the enzyme. The results of these experiments are shown in FIG. 3A. The K_m for glucose-1-phosphate was determined to be 81 μM for the UGPase portion and 87 μM for the PGMase portion of the enzyme.

The conversion of glucose-6-phosphate to UDP-glucose was also measured for the purified enzyme (FIG. 3B). The maximum velocity of UDP-glucose formation (as determined via double reciprocal plots) was 13-fold less for glucose-6-phosphate than for glucose-1-phosphate. The K_m for glucose-6-phosphate in this reaction was 125 μM . These results confirm that the purified UGPase/PGMase protein, which is encoded by the *upp1* gene, is able to catalyze the formation of UDP-glucose from glucose-6-phosphate. A single enzyme has never previously been discovered or purified that is able to catalyze this reaction.

Overexpression Studies

Additional copies of the *upp1* gene were introduced back into *C. cryptica* to demonstrate that the gene confers the predicted activities in vivo. Plasmid pANUP contains a ~6-kilobase *C. cryptica* genomic DNA fragment inserted into the KpnI/EcoRI sites upstream of the *nptIII* gene of plasmid pANRz. pANRz is identical to pACCNPT10 (Dunahay, et al., J. Phycol. 31:1004–1012 (1995)) except that the downstream Bpu102/PstI fragment is replaced by a BglIII/MluI adaptor sequence. The inserted *C. cryptica* frag-

ment contains the entire coding sequence of the *upp1* gene (including introns), as well as 763 bp of noncoding sequence 5' to the coding sequence and approximately 1600 bp of noncoding sequence 3' to the coding sequence. This plasmid was used to transform wild-type *C. cryptica* cells according to the particle bombardment transformation protocol of Dunahay et al. (Dunahay et al., J. Phycol. 31:1004–1012 (1995)). Approximately 5 μg of plasmid was precipitated onto tungsten microcarriers, and 10 μL of the coated particles were used for each bombardment. Plasmid pANRz, which does not contain the *upp1* gene, was used as a control. Bombarded cells were given 2 days to recover before being transferred to selection plates. The transformants were selected based on their resistance to the antibiotic G418 and appeared after about one week. Individual transformants were transferred to liquid 50% ASW medium (Brown, Phycologia 21:408–410 (1982)) and cultured at 26° C. without agitation, under a light intensity of 50 $\mu\text{E}\cdot\text{m}^{-2}\cdot\text{sec}^{-1}$. Cells were harvested for enzyme assays at early stationary phase; 15 mL of each culture were centrifuged at 2000 \times g for 5 minutes. The cell pellet was resuspended in 5 mL of Hepes buffer, pH 7.8, containing 2 mM dithiothreitol, and re-centrifuged. The final cell pellet was resuspended in 0.3 mL of the same buffer. The cell suspension was frozen on dry ice and stored at -80° C. Cell-free extracts were prepared by thawing the frozen cells on ice and centrifuging at 16,000 \times g for 15 min at 4° C. The protein concentrations of the supernatants were measured, and each extract was diluted with the same buffer to protein concentrations of 0.4 and 0.1 mg/mL. UGPase assays were carried out as described in the "Enzyme Assays" section above, using 2 μL of the 0.1 mg protein/mL enzyme extracts. PGMase assays were conducted as described in the "Enzyme Assays" section above, using 50 μL of the 0.4 mg protein/mL enzyme extracts.

The results of assays on 8 transformants are shown in FIG. 4A and FIG. 4B. The activities of both UGPase and PGMase were higher in transformants containing plasmid pANUP than in transformants containing the control plasmid pANRz, indicating that overexpression of the *upp1* gene was achieved. Furthermore, these results suggest that the inserted recombinant *upp1* gene is expressed to produce an enzyme with both UGPase and PGMase activities in vivo.

While various embodiments of the present invention have been described in detail, modifications and adaptations of those embodiments will be apparent to those skilled in the art. It is to be expressly understood, however, that such modifications and adaptations are within the scope of the present invention, as set forth in the following claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(iii) NUMBER OF SEQUENCES: 9

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic oligonucleotide)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

-continued

ARRTRTTNG TRTTRAA

17

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic oligonucleotide)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TGGTAYCCNC CNGGCA

17

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 14 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic oligonucleotide)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GCNGTYTCNA RYTG

14

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 304 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CGGAGACTTG TACGCTGCC TCATCGGCTC AGGTCGTCTC TCTGCCCTCC TCGAAGGAGG 60
 ATACAAGTAC ATGTTCTGCT CAAACTCTGA CAACCTTGGT GCCACCCTTG ACCTGAAAAT 120
 CCTCACCCAC TTCGCCAAAA CGGATGCATC CTTTATGATG GAATGCTGTG AACGCACTGA 180
 AAACGACAAG AAAGGAGGAC ATCTTGTCTG TCGTAATCA GATCAACATT TGATCCTTCG 240
 CGAATCTGCT ATGTGTGCCG ACGAAGACGA GCCTGCCTTC CAAGATATCA CCAAGCACCG 300
 CTTC 304

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 101 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Gly Asp Leu Tyr Ala Ala Leu Ile Gly Ser Gly Arg Leu Ser Ala Leu
 1 5 10 15
 Leu Glu Gly Gly Tyr Lys Tyr Met Phe Val Ser Asn Ser Asp Asn Leu
 20 25 30
 Gly Ala Thr Leu Asp Leu Lys Ile Leu Thr His Phe Ala Lys Thr Asp
 35 40 45
 Ala Ser Phe Met Met Glu Cys Cys Glu Arg Thr Glu Asn Asp Lys Lys

-continued

TCTGACAACC	TTGGTGCCAC	CCTTGACCTG	AAAATCCTCA	CCCAC TTCGC	CAAAACGGAT	1080
GCATCCTTTA	TGATGGAATG	CTGTGAACGC	ACTGAAAACG	ACAAGAAAGG	AGGACATCTT	1140
GCTGTTCGTA	ATTCAGATCA	ACATTTGATC	CTTCGCGAAT	CTGCTATGTG	TGCCGACGAA	1200
GACGAGCCTG	CCTFCCAAGA	TATCACCAAG	CACCGCTTCT	TCAACACGAA	CAATCTTTGG	1260
ATTCGCCTTG	ACAAGTTGCA	GGAGATTGTT	GATAAATATG	GAGGATTCAT	TCCCCTCCCT	1320
ATGATCATGA	ATGCCAAGAC	CGTTGATCCC	AAAGATGACA	ACTCTCAAAA	GGTTCTCCAA	1380
CTCGAAACTG	CTATGGGTGC	TGGTGAGTCC	AACCTTGTA	AAACACTTCA	GTCATTGTGT	1440
TAATACATCA	CTCAGAGGTC	TCTTATATTT	CAAAACAGCC	ATCGAATGCT	TTGATGGTGC	1500
CAGCGCAGTG	GTCGTGCCCT	G TACTCGTTT	CGCCCCGTG	AAGAAGTGCA	ACGACCTTCT	1560
TCTTCTCCGC	AGTGATGCTT	ATGTCATCAC	GGAAGATTTT	CGTCCGGTGC	TCAATCCTCT	1620
TTGCAATGGA	GTTGCCCCCA	TTATTGACCT	GGATTCAAAG	AAGTACAAGC	TAGTCGGATC	1680
TCTGGAGGAA	GCCACAGCCA	ATGGGTGTCC	TTCTCTTGTC	GCTTGTAAGC	GTCTGAAGGT	1740
CAAAGGCACC	ATTCGCTTCG	GCAGATCTAC	CCGTTTGTG	GGAATGTGT	CTATCACTAA	1800
CTCGAGCGAT	GAATCGAAGT	ACGTCTCTGG	AACAATCGAG	AATACAGATC	TCGATGTGTC	1860
TGCTGATACT	GGCCTGGGTC	TTCTCAAGCC	CACTCTTGTC	AGAACTGCGC	CTATTGCAGG	1920
CCAGAAACCT	GGAACCTCCG	GACTTCGCAA	GAAGACTAAG	GAGTTCATGT	CGGAGAACTA	1980
TCTGAGCAAT	TTTGTTCAGT	CTGTATTGCA	TGCTGTCAAT	GCTGCAGGAA	CCAATGTCTC	2040
TGAAGGAACG	CTCATGATTG	GGGGCGACGG	GCGTTATTTT	AACTACTGAGG	CTATTCAAAT	2100
TATCATTAATA	ATGGGTGTTG	CCAACGGGTT	GAAGCGTTTC	TGGATCGGAG	AGAATGGGTT	2160
GCTCTCCACC	CCTGCTGTCT	CTGCCACAAT	TCGAGAGCGT	GGACCAGTAT	GGCAAAAGTC	2220
CTTTGGTGCG	TTCATCCTTA	CCGCAAGTCA	CAACCCTGGA	GGCCCTGAGG	AGGACTTTGG	2280
AATCAAGTAC	AATACCCAGA	ACGGAGGCC	TGCCCCCGAA	TATCTCATGG	AAGCAACTTA	2340
TGCCAACACT	ACTTGCATCA	AAACCTACAA	GATTTGCGAG	GACTTCCCCT	CTATTGACAT	2400
AACTCAAGTC	GGAGCTACCA	CTGTTGCAGC	CGCTGATGGA	AGTACAAGTG	TTGTTGTTGA	2460
GGTGATCCCC	TCGACCCAGT	CTCATGTTAC	CCTACTGAAG	ACCATCTTTG	ACTTCCCTGC	2520
AATCAAGGCG	CCTCTTGACC	GTCCCGATTT	TTCTATGGTC	TACGACTCCA	TGCATGGAGT	2580
TAACGGTCTT	TTTTCCAAGG	CTGTCTTCGT	GGACGAACTT	GGCCAGCCAG	AGTCTGTACT	2640
CAGGAACCAT	ATTCCTAAGG	ACGACTTTGC	CGGTGGACAT	GCTGATCCCA	ACCTTACTTA	2700
CGCCAAAGAG	CTCGTGAAGA	CCATGGGCTT	GGATAGGACT	GGGAACAAGA	TTGATGTTGA	2760
TGGACCCATC	CCTTCTTTTCG	GTGCTGCTGC	TGATGGAGAT	GGTGACCGCA	ACATGATCCT	2820
TGGGACACAG	TTCTTCGTCA	CACCCCTCTGA	TTCTCTAGCT	GTAATTGTTG	CCAATGCCAA	2880
CTGCATCCCA	TTTTTCAGCT	CCCAAGGTGG	TCTCAAGGCC	GTTGCTAGGT	CCATGCCCAC	2940
AAGTGGAGCT	GTAGACCCAG	TAGCAAAGGA	CTTGAACCTT	GACTTCTTTG	AAACACCTAC	3000
TGGATGGAAG	TTTTTCGGCA	ACCTGATGGA	TTCCAAGGCC	ATCTTTAAGG	GCAAAGACTA	3060
CACCCCGTTC	ATCTGCGGTG	AAGAGAGTTT	TGGTACTGGC	TCTGATCATG	TGCGTGAGAA	3120
GGACGGCATA	TGGGCTGTTT	TGGCATGGTT	GAACATCTTG	GCCGCCACA	ACCCTGATGC	3180
ATCGAAGCCT	CTGGTACTGC	TTGAAGACAT	TGTAAGGAAA	CACTGGTCCA	AGTATGGTCG	3240
CAACTACTAC	TGCCGTTGGG	ACTTTGAGGG	TATGGATGCA	GCGGGAGCCA	ATGCCATGAT	3300
GGAGAAGATG	CGCGCTGATG	CAGCATCGAA	CACTGGCCGT	ACTGTTGGAA	GTTACTACTAT	3360
TGCTACTGCC	GATGACTTTA	GGTACGTTGA	CCCCGTGGAT	GGGTCTGTGC	CTGCAAAACA	3420

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AGGAATTCGC TTCCTCATGT CGGATGGATC AAGAGTTATT TTCGTTTGT CGGGCACAGC 3480
GGGTTTCAGGT GCTACTGTTT GCATGTACAT TGAGCAATAT GAAACGGAGA AGCTTGATCT 3540
GCCTGTTGCC TCTGCTCTAG AAGAGCTTAC CTCAATTGCA CTGCAATTAT GTGACATCAA 3600
GACGTTTTGC GGCACCGAAA CTCCAACGT CATCACCTGA 3640

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(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1056 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

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Met Ala Ser Phe Glu Pro Cys Arg Thr Lys Met Glu Ala Glu Gly Ile
 1           5           10          15
Ala Gln Ser Ala Ile Ser Ala Phe Glu Ser Thr Phe Asn Ser Leu Val
          20          25          30
Ser Gly Asn Thr Gly Ile Ile Pro Glu Ser Thr Ile Ser Pro Val Pro
          35          40          45
Glu Leu Val His Thr Asp Ser Ile Thr Ala Glu Pro Asp Ser Ser Leu
          50          55          60
Leu Ala Ser Thr Val Val Leu Lys Leu Asn Gly Gly Leu Gly Thr Gly
          65          70          75          80
Met Gly Leu Asp Lys Ala Lys Ser Leu Leu Glu Val Lys Asn Gly Asp
          85          90          95
Thr Phe Leu Asp Leu Thr Ala Lys Gln Val Met Cys Met Arg Glu Glu
          100         105         110
Phe Gly Gln Lys Val Lys Phe Met Leu Met Asn Ser Phe Ser Thr Ser
          115         120         125
Asp Asp Thr Leu Glu Phe Phe Arg Thr Lys Tyr Pro Thr Leu Ala Ala
          130         135         140
Glu Glu Gly Leu Glu Met Leu Gln Asn Lys Val Pro Lys Ile Asp Ala
          145         150         155         160
Thr Thr Tyr Glu Pro Ala Thr Cys Pro Ser Asp Pro Ser Asn Glu Trp
          165         170         175
Cys Pro Pro Gly His Gly Asp Leu Tyr Ala Ala Leu Ile Gly Ser Gly
          180         185         190
Arg Leu Ser Ala Leu Leu Glu Gly Gly Tyr Lys Tyr Met Phe Val Ser
          195         200         205
Asn Ser Asp Asn Leu Gly Ala Thr Leu Asp Leu Lys Ile Leu Thr His
          210         215         220
Phe Ala Lys Thr Asp Ala Ser Phe Met Met Glu Cys Cys Glu Arg Thr
          225         230         235         240
Glu Asn Asp Lys Lys Gly Gly His Leu Ala Val Arg Asn Ser Asp Gln
          245         250         255
His Leu Ile Leu Arg Glu Ser Ala Met Cys Ala Asp Glu Asp Glu Pro
          260         265         270
Ala Phe Gln Asp Ile Thr Lys His Arg Phe Phe Asn Thr Asn Asn Leu
          275         280         285
Trp Ile Arg Leu Asp Lys Leu Gln Glu Ile Val Asp Lys Tyr Gly Gly
          290         295         300
Phe Ile Pro Leu Pro Met Ile Met Asn Ala Lys Thr Val Asp Pro Lys
          305         310         315         320

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Asp Asp Asn Ser Gln Lys Val Leu Gln Leu Glu Thr Ala Met Gly Ala
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 340 345 350
 Arg Phe Ala Pro Val Lys Lys Cys Asn Asp Leu Leu Leu Leu Arg Ser
 355 360 365
 Asp Ala Tyr Val Ile Thr Glu Asp Phe Arg Pro Val Leu Asn Pro Leu
 370 375 380
 Cys Asn Gly Val Ala Pro Ile Ile Asp Leu Asp Ser Lys Lys Tyr Lys
 385 390 395 400
 Leu Val Gly Ser Leu Glu Glu Ala Thr Ala Asn Gly Cys Pro Ser Leu
 405 410 415
 Val Ala Cys Lys Arg Leu Lys Val Lys Gly Thr Ile Arg Phe Gly Arg
 420 425 430
 Ser Thr Arg Phe Val Gly Asn Val Ser Ile Thr Asn Ser Ser Asp Glu
 435 440 445
 Ser Lys Tyr Val Ser Gly Thr Ile Glu Asn Thr Asp Leu Asp Val Ser
 450 455 460
 Ala Asp Thr Gly Leu Gly Leu Leu Lys Pro Thr Leu Val Arg Thr Ala
 465 470 475 480
 Pro Ile Ala Gly Gln Lys Pro Gly Thr Ser Gly Leu Arg Lys Lys Thr
 485 490 495
 Lys Glu Phe Met Ser Glu Asn Tyr Leu Ser Asn Phe Val Gln Ser Val
 500 505 510
 Phe Asp Ala Val Ile Ala Ala Gly Thr Asn Val Ser Glu Gly Thr Leu
 515 520 525
 Met Ile Gly Gly Asp Gly Arg Tyr Phe Asn Thr Glu Ala Ile Gln Ile
 530 535 540
 Ile Ile Lys Met Gly Val Ala Asn Gly Val Lys Arg Phe Trp Ile Gly
 545 550 555 560
 Glu Asn Gly Leu Leu Ser Thr Pro Ala Val Ser Ala Thr Ile Arg Glu
 565 570 575
 Arg Gly Pro Val Trp Gln Lys Ser Phe Gly Ala Phe Ile Leu Thr Ala
 580 585 590
 Ser His Asn Pro Gly Gly Pro Glu Glu Asp Phe Gly Ile Lys Tyr Asn
 595 600 605
 Thr Gln Asn Gly Gly Pro Ala Pro Glu Tyr Leu Met Glu Ala Thr Tyr
 610 615 620
 Ala Asn Thr Thr Cys Ile Lys Thr Tyr Lys Ile Cys Glu Asp Phe Pro
 625 630 635 640
 Ser Ile Asp Ile Thr Gln Val Gly Ala Thr Thr Val Ala Ala Ala Asp
 645 650 655
 Gly Ser Thr Ser Val Val Val Glu Val Ile Pro Ser Thr Gln Ser His
 660 665 670
 Val Thr Leu Leu Lys Thr Ile Phe Asp Phe Pro Ala Ile Lys Ala Leu
 675 680 685
 Leu Asp Arg Pro Asp Phe Ser Met Val Tyr Asp Ser Met His Gly Val
 690 695 700
 Asn Gly Pro Phe Ser Lys Ala Val Phe Val Asp Glu Leu Gly Gln Pro
 705 710 715 720
 Glu Ser Val Leu Arg Asn His Ile Pro Lys Asp Asp Phe Ala Gly Gly
 725 730 735
 His Ala Asp Pro Asn Leu Thr Tyr Ala Lys Glu Leu Val Lys Thr Met

-continued

740			745			750									
Gly	Leu	Asp	Arg	Thr	Gly	Asn	Lys	Ile	Asp	Val	Asp	Gly	Pro	Ile	Pro
	755						760					765			
Ser	Phe	Gly	Ala	Ala	Ala	Asp	Gly	Asp	Gly	Asp	Arg	Asn	Met	Ile	Leu
	770						775					780			
Gly	Thr	Gln	Phe	Phe	Val	Thr	Pro	Ser	Asp	Ser	Leu	Ala	Val	Ile	Val
	785				790						795			800	
Ala	Asn	Ala	Asn	Cys	Ile	Pro	Phe	Phe	Ser	Ser	Gln	Gly	Gly	Leu	Lys
			805								810			815	
Ala	Val	Ala	Arg	Ser	Met	Pro	Thr	Ser	Gly	Ala	Val	Asp	Arg	Val	Ala
			820					825					830		
Lys	Asp	Leu	Asn	Leu	Asp	Phe	Phe	Glu	Thr	Pro	Thr	Gly	Trp	Lys	Phe
	835						840					845			
Phe	Gly	Asn	Leu	Met	Asp	Ser	Lys	Ala	Ile	Phe	Lys	Gly	Lys	Asp	Tyr
	850						855				860				
Thr	Pro	Phe	Ile	Cys	Gly	Glu	Glu	Ser	Phe	Gly	Thr	Gly	Ser	Asp	His
	865				870					875					880
Val	Arg	Glu	Lys	Asp	Gly	Ile	Trp	Ala	Val	Leu	Ala	Trp	Leu	Asn	Ile
			885						890					895	
Leu	Ala	Ala	His	Asn	Pro	Asp	Ala	Ser	Lys	Pro	Leu	Val	Thr	Val	Glu
			900					905					910		
Asp	Ile	Val	Arg	Lys	His	Trp	Ser	Lys	Tyr	Gly	Arg	Asn	Tyr	Tyr	Cys
	915						920					925			
Arg	Trp	Asp	Phe	Glu	Gly	Met	Asp	Ala	Ala	Gly	Ala	Asn	Ala	Met	Met
	930					935					940				
Glu	Lys	Met	Arg	Ala	Asp	Ala	Ala	Ser	Asn	Thr	Gly	Arg	Thr	Val	Gly
	945				950					955					960
Ser	Tyr	Thr	Ile	Ala	Thr	Ala	Asp	Asp	Phe	Arg	Tyr	Val	Asp	Pro	Val
			965						970					975	
Asp	Gly	Ser	Val	Ala	Ala	Lys	Gln	Gly	Ile	Arg	Phe	Leu	Met	Ser	Asp
			980					985					990		
Gly	Ser	Arg	Val	Ile	Phe	Arg	Leu	Ser	Gly	Thr	Ala	Gly	Ser	Gly	Ala
		995					1000				1005				
Thr	Val	Arg	Met	Tyr	Ile	Glu	Gln	Tyr	Glu	Thr	Glu	Lys	Leu	Asp	Leu
	1010					1015				1020					1
Pro	Val	Ala	Ser	Ala	Leu	Glu	Glu	Leu	Thr	Ser	Ile	Ala	Leu	Gln	Leu
	025				1030					1035				1040	
Cys	Asp	Ile	Lys	Thr	Phe	Cys	Gly	Thr	Glu	Thr	Pro	Thr	Val	Ile	Thr
			1045					1050					1055		

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 9 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Phe Met Leu Met Asn Ser Phe Ser Thr
 1 5

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 amino acids

-continued

(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Ala Val Phe Val Asp Glu Leu Gly Gln Pro Glu Ser Val Leu Arg Asn
1 5 10 15

His Ile Pro

What is claimed is:

1. An isolated and purified DNA encoding a protein from *Cyclotella cryptica* comprising an enzyme selected from the group consisting of:
 - (a) UDP-glucose pyrophosphorylase;
 - (b) phosphoglucomutase; and
 - (c) UDP-glucose pyrophosphorylase and phosphoglucomutase.
2. The DNA of claim 1 wherein the selected enzyme are UDP-glucose pyrophosphorylase and phosphoglucomutase.
3. The DNA according to claim 1 wherein the amino acid sequence of the encoded protein is the sequence identified as SEQ ID NO:7.
4. A vector comprising the DNA of claim 1.
5. A vector comprising the DNA of claim 3.
6. A host containing the vector of claim 4.
7. A host containing the vector of claim 5.
8. A host containing the vector of claim 5, wherein said host is *Cyclotella cryptica*.
9. An isolated and purified DNA having the sequence identified as SEQ ID NO:6.
10. The DNA of claim 9 wherein the DNA encodes UDP-glucose pyrophosphorylase and phosphoglucomutase.
11. An isolated and purified DNA from *Cyclotella cryptica* encoding UDP-glucose pyrophosphorylase and phosphoglucomutase.

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