

Volumes 113–116 (12 Issues), Spring 2004, ISSN: 0273–2289

Applied Biochemistry and Biotechnology

Executive Editor: Ashok Mulchandani

Biotechnology for Fuels and Chemicals

The Twenty-Fifth Symposium

Editors

Mark Finkelstein

James D. McMillan

Brian H. Davison

Barbara Evans

 **HUMANA PRESS**

Plant Biotechnology and Feedstock Genomics

JAMES S. MCLAREN¹ AND STEVEN R. THOMAS²

¹*StrathKirn Inc., Chesterfield, MO; and*

²*National Renewable Energy Laboratory, Golden, CO*

Over the first part of the twentieth century, it is expected that a large increase in the utilization of renewable bioresources will occur. The possibility for change arises from the broad range of research projects that are currently under way, and the drivers for change continue to include sustainable economic growth and national energy security, and to minimize anthropogenic effects on the environment.

The recent research focus on conversion of conventional lignocellulose biomass will provide useful results in clarifying potential application within various processing mills. However, additional technical approaches will also contribute to successful implementation of the overall biomass platform. For example, if the raw input material were to be “designed” to be more easily converted to a sugar platform, then the system efficiency, practical application, and economic viability would all increase. The recent significant advances in genomics and biotechnology applications provide a new opportunity to utilize renewable resources in ways that will help ensure sustainable enterprises.

The knowledge and understanding of genome sequences, gene function, gene expression, protein interactions, and metabolic control mechanisms will form the basis for future biotech-based alterations in primary production, which is the key underlying renewable resource. The participants in this session provided an exciting view, covering state-of-the-art developments in several relevant areas. Details are provided in the individual papers and an overview of the session is as follows:

Nathan Lakey (Orion Genomics) introduced the exciting capability to uncover the sequences of genes within the genome, without the need to sequence all the nongene DNA space. The technique takes advantage of a chemical difference between repetitive junk DNA, which is methylated and comprises ~90% of plant genomes, and genes, which remain unmethylated and occupy only 10% of the genome. This approach has been demonstrated in all of the major plant species and is estimated to provide the “genespace” sequence at a 10-fold lower cost and much faster than previous methods. The ultimate application to plants used for bioenergy would provide an operational base for significant improvements, primary

productivity, and composition, leading to much lower cost of input “sugars” and easier bioconversion to intermediates of interest.

Mike Lassner (Verdia Inc.) presented examples of the usefulness of directed molecular evolution as an *in vitro* process that more easily achieves what was traditionally attempted via reproductive crossing and recurrent selection (plant breeding). Proteins may be engineered that have specific desirable characteristics via methods that “evolve” the basic underlying DNA. For example, the outcome can be enzymes with improved kinetic properties that result in enhanced primary production, or proteins that remain operational under extreme conditions. In addition, compositional proteins may be enhanced to provide functional performance that was not achievable via conventional methods.

Justin Stege (Diversa Corporation) discussed the molecular evolution of enzymes for particular pathways, with a focus on the modification of oil composition. Oleochemical applications for such enzymes include applications as biocatalysts for fatty acid modifications. In a program to integrate production and processing, such enzymes can be used to modify the fatty acid content of vegetable oils *in planta*. Results show that expressing such new enzymes in oilseed crops has resulted in altered oil composition, and that the features may be used to better design plant-based oils for use as biofuels and as improved renewable feedstocks.

Mariam Sticklen (Michigan State University) presented results showing the successful expression of three different full-length polyhydroxybutyrate (*phb*) genes in maize, and accumulation of the poly-(*R*)-3-hydroxybutyrate (PHB) enzymes within maize chloroplasts. PHB is a potentially useful polymer for use in the plastic markets and is currently produced via fermentation. PHB from transgenic plants was first demonstrated using *Arabidopsis*, but expression in major crops has added economic implications. This paper also reported work on codon-optimized *Trichoderma reesei cbh1* and wild-type *Acidothermus cellulolyticus e1* genes regulated by the rice *rbcs* promoter. Many maize plants were transformed with evaluations of protein targeting to various subcellular organelles.

Steve Thomas (National Renewable Energy Laboratory) described the use of near-infrared (350–2500 nm) reflectance spectroscopy as a genetic screening tool to identify individual plants with particular cell-wall compositions. A broad range of corn genotypes was evaluated, including mutant populations, and the chemical composition was aligned with the phenotypic characteristics. Quantification of plant cell walls is an essential component if lignocellulose biomass is to become a major contributor to a biobased economy. Through such measurements it will be possible to better match biocatalysis to input composition, and to enhance raw material inputs through modified composition with more desirable characteristics.

Deborah Samac (USDA-ARS, Minnesota) reported that more than 100 genes are involved in cell-wall biosynthesis, and that many of these genes have now been cloned to better understand how modification could be achieved. Experiments on alfalfa involving the expression of a soybean

UDP-glucose dehydrogenase cDNA under the control of two promoters active in vascular tissues showed good expression in greenhouse tests. Other results provided evidence that altering the expression of a single gene may have only minor effects. Multigene changes or finding specific regulatory genes may be required to improve the ability to modify plant cell walls to have more useful features.

Overall the participants in this session covered a wide range of topics and clearly demonstrated the importance of genomics and biotechnology to the future of renewable resources. The major underlying threads seemed to be the following:

1. Rapid advances in the technology are opening the door for novel opportunities.
2. Enzymes can be improved via molecular evolution, and for use *in planta*.
3. Single gene changes may not be enough for coordinated pathway engineering.
4. Compositional quantification is important for baseline and useful modifications.

Above all, there was general agreement that the integration of these approaches may be much more productive than just exploring limited single areas. The true biorefinery concept requires integration of all the available science—to enable a broad range of input types along with an output portfolio that includes biofuels and a selected range of economically acceptable biobased products.

National Renewable Energy Laboratory
PERMISSION TO USE COPYRIGHTED MATERIAL

PERMISSIONS

I give the National Renewable Energy Laboratory (NREL) of Golden, Colorado, irrevocable and worldwide permission to post the following onto its site on the World Wide Web (Web).

- Yes
 No

Name of document, report, article, etc.: Eight U.S. Department of Energy Biomass Program (NREL, ORNL, ANL, INEEL, PNNL) authored articles from 25th Symposium on Biotechnology for Fuels and Chemicals, Breckenridge, CO, May 2003 — Applied Biochemistry and Biotechnology, Volumes 113-116, Spring 2004, ISSN 0273-2289 pp. 13-26, 113-114, 653-670, 807-826, 871-886, 1127-1138, 1139-1162, 1163-1168.

In addition to posting on the Web, I give NREL worldwide permission to distribute hard copies of the document in response to requests.

- Yes
 No

*****PLEASE NOTE*****

- For jointly authored works, only one signature is required, but we assume all authors have been advised and have consented to the terms of this form.
- If you are employed and you prepared your work as a part of your job, the rights to your work initially rest with your employer. In that case, when you sign this form, we assume you are authorized to do so by your employer and that your employer has consented to the terms and conditions of this form. If not, it should be signed by someone so authorized.

When posting on the Web, NREL will credit the copyright holder thus: **Posted on this site with permission from Humana Press**

When distributing hard copies, NREL will label the document: **Reprinted with permission from Humana Press**

I was a U.S. government employee when I produced this work as part of my federal employment and therefore the work is not subject to U.S. copyright protection.

I deny permission to post the document on the Web or to distribute hard copies, but give permission for the document to be abstracted and cited in NREL's database, along with information about obtaining the document directly from the copyright holder.

The document can be obtained by writing:

SIGNATURE

I represent and warrant that I have the full right and authority to grant the rights herein granted, that no one else's permission is required, and that should my work contain previously published and/or copyrighted material that permission has been obtained for such use and that any required credit lines and copyright notices are duly noted.

Thomas B Lamigan
Printed name

Vice President, Humana Press
Title and Company

Thomas B Lamigan
Signature

5/27/04
Date