Biotechnology & Biomaterials

Chou et al., J Biotechnol Biomater 2015, 5:2
http://dx.doi.org/10.4172/2155-952X.1000179

Research Article Open Access

Genetic Engineering and Improvement of a *Zymomonas mobilis* for Arabinose Utilization and Its Performance on Pretreated Corn Stover Hydrolyzate

Yat-Chen Chou*, Jeffrey Linger, Shihui Yang and Min Zhang

National Bioenergy Center, National Renewable Energy Laboratory, Golden, CO 80401, USA

Abstract

A glucose, xylose and arabinose utilizing *Zymomonas mobilis* strain was constructed by incorporating arabinose catabolic pathway genes, *araBAD* encoding L-ribulokinase, L-arabinose isomerase and L-ribulose-5-phosphate-4-epimerase in a glucose, xylose co-fermenting host, 8b, using a transposition integration approach. Further improvement on this arabinose-capable integrant, 33C was achieved by applying a second transposition to create a genomic knockout (KO) mutant library. Using arabinose as a sole carbon source and a selection pressure, the KO library was subjected to a growth-enrichment process involving continuous sub-culturing for over 120 generations. Strain 13-1-17, isolated from such process demonstrated significant improvement in metabolizing arabinose in a dilute acid pretreated, saccharified corn stover slurry. Through Next Generation Sequencing (NGS) analysis, integration sites of the transposons were identified. Furthermore, multiple additional point mutations (SNPs: Single Nucleotide Polymorphisms) were discovered in 13-1-17, affecting genes *araB* and *rpiB* in the genome. We speculate that these mutations may have impacted the expression of the enzymes coded by these genes, ribulokinase and ribose 5-P-isomerase, thus attributing to the improvement of the arabinose utilization.

Keywords: *Zymomonas mobilis*; Arabinose; *araBAD*; Transposition; Knockout library; Cre-Lox recombination; Next generation sequencing (NGS)

Introduction

Z. mobilis has been recognized as a viable alternative to yeast for ethanol production from biomass for its high conversion yield, high ethanol tolerance, resistance to pretreated hardwood hydrolyzate and fermentation capability at lower pH and in anaerobic conditions [1]. However, the narrow substrate utilization, capable of only using glucose, fructose and sucrose, hinders the application of this organism in the ethanol production from lignocellulosic biomass feedstock. Our laboratory has successfully constructed a glucose/xylose co-utilizing strain, *Z. mobilis* 8b, based on ZM4 (ATCC31821), using homologous recombination and transposition integration methods [2]. Moreover, strain 8b demonstrates superior ethanol production performance in a pretreated corn stover hydrolyzate [3].

Within the context of process economics for cellulosic ethanol, it is imperative to maximize sugar utilization. Given that arabinose comprises approximately 5% of the available sugars within corn stover [4], any organism that is unable to utilize this sugar will automatically leave valuable sugars (and potential end-product) behind. Specifically, NREL's 2011 design report for biochemical conversion of lignocellulosic biomass to ethanol [4] assumes an 85% conversion of arabinose to ethanol in order to meet the aggressive 2012 cost targets of $2.15/gal in year 2007 dollars. Importantly, achieving this 85% conversion reduces the MESP by $0.08/gal. Therefore, we sought to generate an efficient arabinose utilizing strain of *Z. mobilis*.

Previous genetic engineering work has implemented arabinose utilization capabilities within *Z. mobilis* strain ATCC39676 [5]. However, its arabinose usage and coordinate ethanol production were limited. We hypothesized that part of this could be due to the strain background used, as ATCC39676 generally seems to be inferior to strains based on ZM4 with regards to ethanol production. For example ZM4-based glucose/xylose co-utilizing, plasmid-bearing strains have proven superior as it relates to ethanol production from oat hull hydrolyzate [6] and resistance to acetic acid inhibition [3,6] as well as elevated temperature [3]. More recently, DuPont has also constructed an arabinose/xylose co-utilizing strain based on the ZM4 host and improved its arabinose utilization by the introduction of an arabinose-proton symporter [7].

We report here the generation of arabinose-utilizing strains of *Z. mobilis* with superior performance in lab pure sugar media as well as a corn stover derived biomass. These strains were generated through the use of rational genetic engineering, transposon based mutagenesis, and selection/enrichment in arabinose medium. We show these strains are superior not only in the usage of arabinose in defined rich media, but also in process relevant conditions using saccharified pre-treated corn stover as the nutrients source when compared with their parental strain, 8b introduced with an arabinose pathway genes (namely, Strain 33C). We further define the underlying genetic alterations that have occurred within these strains and hypothesize about their roles in increasing arabinose utilization for future metabolic engineering to further enhance arabinose utilization.

Materials and Methods

Bacterial strains and growth conditions

All bacteria and plasmids used in this study are listed in Table 1. Escherichia coli DH5α and SCS110 were cultured in LB medium

***Corresponding author:** Yat-Chen Chou, National Bioenergy Center, National Renewable Energy Laboratory, 15013 Denver West Parkway Golden, CO 80401, USA, Tel: 3033847766; E-mail: yat.chen.chou@nrel.gov

Received March 18, 2015; **Accepted** April 21, 2015; **Published** April 28, 2015

Citation: Chou YC, Linger J, Yang S, Zhang M (2015) Genetic Engineering and Improvement of a Zymomonas mobilis for Arabinose Utilization and Its Performance on Pretreated Corn Stover Hydrolyzate. J Biotechnol Biomater 5: 179. doi:http://dx.doi.org/10.4172/2155-952X.1000179

Copyright: © 2015 Chou YC, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Strain or plasmid		Description	Reference
Strain			
Escherichia coli	DH5α	F– Φ80*lac*ZΔM15 Δ(*lac*ZYA-*arg*F) U169 *rec*A1 *end*A1 *hsd*R17 (rK–,mK+) *pho*A *sup*E44 λ– *thi*-1 *gyr*A96 *rel*A1	Invitrogen
	SCS110	*rpsL* (Str-r) *thr leu endA thi-1 lacY galK galT ara tonA tsx dam dcm supE44 Δ(lac-proAB) [F' traD36 proAB lacIqZΔM15]*	Agilent Technologies
Zymomonas mobilis	8b	ZM4 integrated with Tc-P_{gap}*xylAB*/Cm-*Penotaltkt*; glucose/xylose utilizing	our lab; [2]
	Strain 33	8b integrated with *lox*P-*aadA-loxP-*P_{gap}*-araBAD;* glucose/xylose/arabinose utilizing	this study
	33C	*aadA* free derivative of Strain 33; glucose/xylose/arabinose utilizing	this study
	13-1-17	isolate from 33C genomic KO library; glucose/xylose/arabinose utilizing	this study
	15-1-34	isolate from 33C genomic KO library; glucose/xylose/arabinose utilizing	this study
Plasmid	pMOD2Sp	Amp^r,Sp^r; integrative plasmid containing ME-*loxP-aadA-loxP*-ME for transposition and KO library construction; non-replicative in *Z. mobilis*	this study
	pMOD2Sploxara	Amp^r,Sp^r; integrative plasmid containing ME-*loxP-aadA-loxP-*P_{gap}*araBAD*-ME for transposition; non-replicative in *Z. mobilis*	this study

Table 1: Bacterial strains and plasmids.

at 37°C with shaking (200 rpm) and *Z. mobilis* strains were cultured microaerophilically (either statically or with gentle shaking) in rich media ("RM": 10 g/L yeast extract, 2 g/L KH_2PO_4) supplemented with glucose or arabinose ("RMG" or "RMA," respectively). For regeneration and selection of *Z. mobilis* transformants or integrants, mating medium ("MMG"; 10 g/L yeast extract, 5 g/L tryptone, 2.5 g/L $(NH_4)_2SO_4$, 0.2 g/L K_2HPO_4 and 50 g/L glucose) was used. All agar media were made with 15 g/L agar. Appropriate antibiotics were added to the media when needed: Spectinomycin ("Sp", 200 μg/mL for *Z. mobilis* and *E. coli*), ampicillin ("Amp", 100 μg/mL) and kanamycin ("Kan", 50 μg/mL).

Recombinant DNA techniques

Plasmid DNA isolation, restriction endonuclease digestion, ligation and transformation, agarose electrophoresis and other standard recombinant DNA techniques were carried out according to the manufacturers' recommendations. Plasmid DNA extraction was carried out using Qiaprep Spin Miniprep kits and gel extraction with Qiaquick kits (Qiagen, CA).

Construction of plasmids and linear DNA for transposition integration

An EZ-Tn5™ transposon construction vector (non-replicative in Zymomonas), pMOD2 (Epicentre, WI) was used to construct plasmids for the introduction of arabinose catabolic pathway genes or transposition mutagenesis. Mosaic ends (ME) flanking a multiple cloning sites in pMOD2 are designed for the cloning and production of transposons. Plasmid pMOD2Sploxara (Figure 1A) was assembled by cloning in *Sbf*I site of pMOD2 a DNA fragment containing two elements: 1) P_{gap}-*araBAD* operon [5], and 2) *aadA* gene (conferring spectinomycin resistance; GI:125380577/DQ975275.1) [8] flanked by *loxP* sites (*loxP-aadA-loxP*). The *loxP* sites were used in conjunction with a plasmid-borne copy of Cre recombinase for the removal of *aadA* from the genomes of integrants when required. Plasmid pMOD2Splox (Figure 1B) was constructed by *NotI* digestion of pMOD2Sploxara followed by self-ligation to remove the P_{gap}-*araBAD*.

Transposition integration of *loxP-aadA-loxP-*P_{gap}*-araBAD* or *ME-loxP-aadA-loxP-ME* in *Z. mobilis* 8b and removal of *aadA* in the integrants

A commercially available system, EZ::TN™ Transposome™ Kit (Epicentre, WI) [9] was used to generate transposomes (a DNA-transposase complex or specifically "transposase bound ME-*loxP-aadA-loxP-*P_{gap}*-araBAD*-ME or ME-*loxP-aadA-loxP*-ME") for initiating the transposition in *Z. mobilis*. Integrative plasmid, pMOD2Sploxara, or *Pvu*II-digested ME-*loxP-aadA-loxP*-ME (from pMOD2Splox) was treated with transposase at room temperature for one hour followed by dialysis against 5% glycerol and 5 mM Tris-HCl (pH7.8) before electroporation/transformation in *Z. mobilis* 8b [2]. A cre-loxP site-specific recombination [10] was used to remove the *aadA* from the integration site in the genome by expressing a Cre recombinase gene (*cre*) on a plasmid in the integrants.

Transformation of *Z. mobilis*

Z. mobilis or *E.coli* cells were transformed with plasmids by electroporation (Bio-Rad Gene Pulser, 0.1-cm gap cuvettes, 1.6 kV, 200 ohms, 25 μFD). Electrocompetent *Z. mobilis* or *E. coli* cells were prepared by collecting cells at an optical density (OD_{600}) = 0.4 to 0.6, washed once in ice-cold sterile water followed by 10% glycerol and concentrated approximately 1000-fold. Competent cells were stored at –80°C as small aliquots for later use. Transformants of *E. coli* or *Z. mobilis* were selected on LB or MMG, agar plates, respectively, containing antibiotics. Due to the presence of a restriction/modification system, all plasmids were built in and isolated from a methylation-deficient *E. coli* strain, SCS110, for successful transformations in *Z. mobilis* ZM4 derived hosts, 8b or 33C.

Genomic Knockout (KO) library construction based on strain 33C using the transposition integration method

An *aadA*-free, arabinose utilizing strain, 33C, was further used to construct a genomic library with non-specific insertion of the *aadA* gene by transposition. A transposome (*Pvu*II digested ME-*loxP-aadA-loxP*-ME fragment of pMOD2Splox, bound by transposases) was used to transform strain 33C by electroporation. Colonies that appeared on MMG plates with Sp were collected by scraping and rinsing the plates with a small volume of RM with 5% glucose and Sp. Collected cells were added with 1/3 volume of the same medium and incubated at 33°C for 1 h before aliquoting and freezing in the presence of 20% glycerol. Multiple transpositions and transformations were conducted in order to obtain sufficient colonies to represent the coverage of a *Z. mobilis* genome.

Enrichment of 33C knockout (KO) library in medium containing arabinose

Frozen library cells were thawed and revived for 2 h at 35°C in 20-mL modified rich medium containing 10 g/L yeast extract, 2 g/L KH_2PO_4, 7.5 g/L glucose, 2.5 g/L L-arabinose, 1g/L $MgSO_4$ and Sp200. The revived cultures were used to inoculate 100-mL the same

medium containing different sugar composition (5 g/L glucose and 5 g/L L-arabinose) in shake flasks as seeds. Seeds were incubated at 35°C overnight before inoculating RMA5 (50 g/L L-arabinose, initial OD600 = 0.05) for the enrichment process. Growth was monitored by OD measurement. RMA5 grown cultures at mid to late exponential phase were diluted into fresh RMA5 daily (or every 2-3 days, dependent upon the growth rates) to enrich for the fast growers in arabinose. As a non-mutated control, 33C was subjected to the same enrichment process. Periodically, cultures were streaked on RMA5 agar plates for single colony isolation. Large colonies were selected and grown in RMA5 and preserved at -80°C for later evaluation.

Evaluation of arabinose utilizing *Z. mobilis* in mixed sugar medium in shake flasks

For down selection of the fast arabinose growers from the KO library, frozen stocks of single isolates from the enrichment process were evaluated in shake flasks containing RM supplemented with sugars – 1% glucose; 4% xylose; 1% arabinose (RMG1X4A1). Frozen stocks were inoculated in 3-mL RMG2 (2% glucose) for 6 hours at 33°C, then diluted into 20-mL RMG6 (6% glucose) as seeds and grown at 33°C for 16 hours with shaking. Seed cultures were concentrated by centrifugation before inoculating the mixed sugar medium (RMG1X4A1) in shake flasks (initial OD = 0.05). Fermentation was conducted at 33°C with 100 rpm shaking. The initial pH of the medium was 6.0 but was not controlled or monitored during the fermentation.

Pretreated, saccharified corn stover slurry ("PCS Saccharified Slurry") production

For slurry A, the corn stover feedstock (with 34% total solids content) used in this study was knife-milled and de-acetylated using 0.4% NaOH (w/w) at 80°C followed by a dilute-acid pretreatment (0.5% H_2SO_4, 150°C, 20 minutes). Pretreated corn stover (PCS) was neutralized to pH 5.1 using 30% ammonium hydroxide. Enzymatic hydrolysis was performed using Novozymes' Cellic CTec 2 at a loading of 40 mg protein/g cellulose at 48°C for 96 hours. Slurry B was produced without the de-acetylation step. For fermentation purposes, the slurries were supplemented with RM and diluted with water to an equivalence of 20% (for slurry A) or 15% (for slurry B) total solids (TS) in the original corn stover feedstock before the inoculation of cultures.

Evaluation of arabinose utilizing *Z. mobilis* in the saccharified slurries (20% and 15% total solids for slurries A and B, respectively)

Two strains selected from the mixed sugars fermentation were evaluated in fermentors (Sixfor) containing PCS saccharified slurry at 20% total solids (TS) for slurry A and 15% TS for slurry B. Frozen cultures were revived in 4-mL RMA5 or RMG5 for 15-16 h followed by inoculation in 20-mL RMG5 for 6 h as preseeds. Seeds were prepared by diluting the preseeds in 100-mL seed medium (RM with 10% glucose and 2% xylose) supplemented with 1 g/L sorbitol and grown for 17 -18 h before inoculating the fermentors. Fermentors were filled with 180 mL of saccharified slurry (containing RM), to which 20-mL seeds were added, bringing the TS concentration to approximately 18% (slurry A) and 13.5% (slurry B) for the fermentations. Temperature was maintained at 33°C and pH 5.8 with KOH. Initial OD was 1.0. Glucose, xylose and arabinose concentrations were measured by high performance liquid chromatography (HPLC) using an Agilent 1100 series HPLC (Santa Clara, CA) with a carbohydrate column (Shodex SP0810) at 85°C and a Bio-rad de-ashing guard p/n 125-0118. Mobile phase used was water and flow rate at 1 mL/min. Ethanol, acids and furfural were measured by the HPLC equipped with an organic acid column (Bio-rad HPX-87H) at 55°C and a Bio-rad H+guard p/n 125-0129. Sulfuric acid (0.01N) was used as the mobile phase, at a flow rate of 0.6 mL/min. Refractive Index detector (RID) was used in both HPLC analyses.

Genomic resequencing through next-generation sequencing (NGS)

Genomic DNA was extracted using Qiagen DNAEasy Kit and sent to the University of Utah for next-generation sequencing using Illumina Hiseq2000 after passing the quality requirements. The quality of FASTQ genome resequencing data was checked using FastQC program (http://www.bioinformatics.babraham.ac.uk/projects/fastqc/). Data passing the quality control was imported into CLC Genomics Workbench (version 5.1, CLCBio, Denmark) for data analysis to identity the potential small genetic changes such as SNPs (Single Nucleotide Polymorphisms) and Indels (Insertions/Deletions). Briefly, sequencing reads were trimmed to remove reads with poor quality and then mapped against the reference genome sequence followed by resequencing analysis using both Probabilistic Variant Detection and Quality-based Variant Detection tools. The variants identified were filtered to remove those with less than 80% confident frequency level [11] before comparing different strains. In addition, the genomic loci of transposon insertion were determined through identifying the reads containing the transposon sequences and subsequently Blasting (BlastN, NCBI) the flanking sequences (of the transposon) against the reference genome, *Z. mobilis* ZM4 [12].

Results

Introduction of arabinose catabolic pathway in *Z. mobilis* 8b

To create a glucose/xylose/arabinose utilizing *Z. mobilis* strain, arabinose catabolic pathway genes from *E. coli* (*araBAD*, a three-gene operon encoding L-ribulokinase, L-arabinose isomerase and L-ribulose-5-phosphate-4-epimerase) were introduced in the genome of a glucose/xylose utilizing host 8b [2] adopting a transposition method with a non-replicative plasmid pMOD2Sploxara (Figure 1A) and transposase. A native promoter of glyceraldehyde-3-phosphate dehydrogenase gene (ZMO0177;"P_{gap}"), isolated from *Z. mobilis* ZM4 was used to drive the expression of *araBAD* [5]. Antibiotic marker gene, *aadA* was included in the integration fragment for the selection of transformants/integrants. Multiple Sp^r integrants were isolated and subjected to screening processes in RMA medium (2% arabinose) in test tubes. In this initial screening/selection process, integrants exhibiting incremental improvement in growth rates when serially cultured in the same medium (Supplementary, Figure S1) were further used for single colony isolation on RMA plates (5% arabinose). One colony, namely "Strain 33" demonstrated the highest arabinose utilization in a preliminary shake flask evaluation in RMG0.8X4.5A0.6 (0.8%glucose, 4.5%xylose and 0.6% arabinose) (Supplementary, Figure S2). While Strain displayed the ability to utilize arabinose, the efficiency was lower than desired. Thus, we chose to use this strain as the base "chassis" for further genetic manipulation to improve the arabinose conversion efficiency. Due to the very limited antibiotic markers that are available to *Zymomonas* in general, we first created an *aadA* free derivative of Strain 33 taking advantage of the Cre-Lox recombination system. Two copies of short *loxP* sequence (flanking *aadA*) were built in the integration cassette for this very purpose. Marker gene, *aadA* was removed from the integration cassette, *aadA*- P_{gap}-*araBAD*, leaving only the P_{gap}-*araBAD* in the genome of Strain 33. The resultant *aadA*-free, arabinose-utilizing derivative was named 33C. A mixed sugar (glucose, xylose and arabinose) fermentation in shake flasks was conducted to

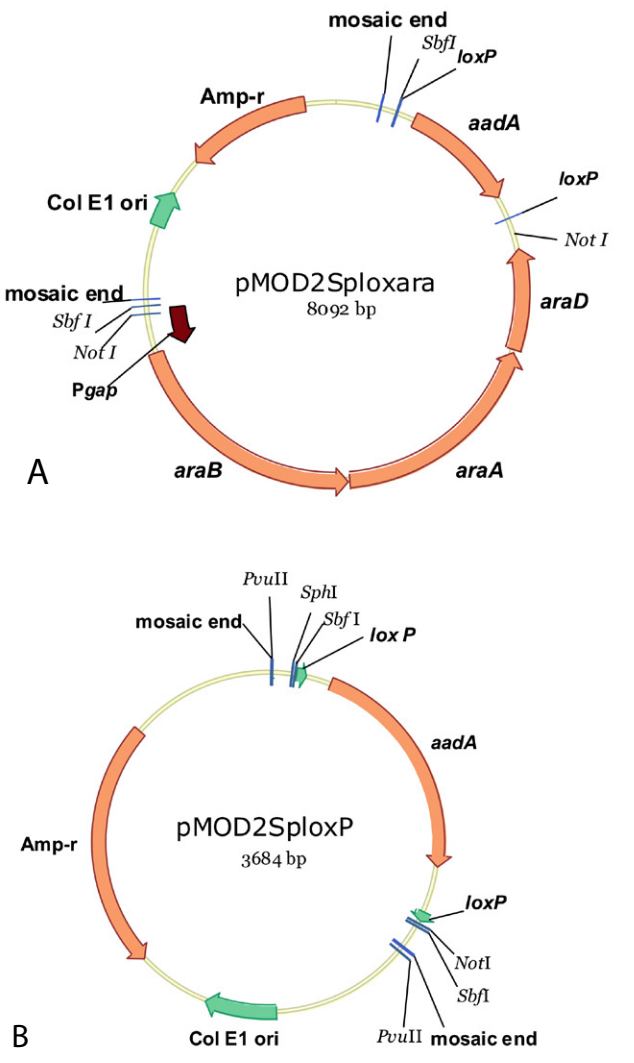

Figure 1: Plasmid maps of pMOD2Sploxara (A) and pMOD2SploxP (B).

compare the performance of strains 33 and 33C. Results confirmed the latter strain maintained the same fermentation capability as its parent pertaining to sugar utilization and ethanol yield (Supplementary, Figure S3).

Knockout library construction based on 33C and the selection/enrichment in arabinose medium

It has been reported that adaptation improved sugar utilization in *Zymomonas* [2,13] and *Pseudomonas* [14] after rational metabolic engineering. Therefore, we additionally reasoned that following implementation of arabinose metabolic genes, other un-specified genetic changes might be required to achieve optimal arabinose metabolism. The simple incorporation of heterologous genes might render native metabolic networks out of balance. Accordingly, we proposed using transposon based mutagenic method, which maximizes the diversity of genomic loci for the insertion (of an antibiotic marker gene) to potentially mutagenize any genes or regulatory elements in the genome. This library of mutants are then collectively subjected to continuous growth in the presence of arabinose to encourage the selection/enrichment of superior arabinose utilizing strains of *Z. mobilis* as compared with previous generations.

A second transposition process was deployed to the *aadA*-free, arabinose utilizing strain, 33C for the construction of a genomic knockout (KO) library to further improve on arabinose growth. This mutant library was generated by non-specific insertions of a transposome cassette, *Pvu*II digested fragments (ME-*loxP*-*aadA*-*loxP*-ME) treated with transposases, in the genome of 33C by electroporation. Nearly 20,000 Spr colonies, representing over eight-fold coverage of *Z. mobilis* genome, were collected and preserved at -80°C as the library. To select and enrich for the fast growers in arabinose medium, library cells were revived from frozen stocks (KO1 and KO2, two individual vials serving as replicates) and inoculated in rich medium with arabinose as the sole carbon source (RMA5) at 35°C. Cultures at exponential growth phase were transferred into fresh RMA5 for 17 consecutive transfers. Figure 2 shows representative examples of the growth curves from these transfers. 33C was grown and transferred alongside the library cultures as a control. This was done to understand the rate of adaptation in homogeneous parent culture, as compared to our heterogeneous transposon disruption library.

Library cultures showed higher final cell density and faster growth rate than the control starting from 12th transfer, suggesting an improved strain(s) had populated and become dominant in the culture. Further transfers showed stabilized growth patterns between library cultures and control 33C. Single colonies were isolated from the 5th to 17th transfers on RMA5 plates. Representative large colonies from each transfer were evaluated in a pure sugar medium (RMG1X4A1) without pH control for the purpose of down-selection. Based on arabinose utilization, two colonies (13-1-17 and 15-1-34, from 13th and 15th transfers respectively) exhibiting higher arabinose utilization than 33C and the rest of the isolates (Figure 3), were chosen for evaluation in a PCS saccharified slurry. Strain 16-3-30 though relatively high arabinose utilization, was not included in the hydrolyzate evaluation because of the lower ethanol yield (data not shown).

Fermentation performance of select library cultures in corn stover hydrolyzates

While pure sugar fermentation indicated promising results by 13-1-17 and 15-1-34, it is crucial that the strains can perform in a less favorable environment (yet closer to the industrial operation application), i.e. hydrolyzates produced through dilute-acid pretreatment, which is known to generate compounds that are inhibitory to microorganisms [15]. Strains 13-1-17 and 15-1-34, demonstrating improvement of arabinose utilization in a pure sugar medium, were evaluated in a pH-controlled fermentation with PCS saccharified slurries (Slurry A: 20% or Slurry B: 15% total solids, pretreated and enzymatically hydrolyzed). Due to the limited quantity of each slurry, we were not able to incorporate duplicates of each strain using the same slurry. Rather, we used two different slurries as substrates in the two independent fermentation experiments to show the consistent improvement of strains 13-1-17 (and 15-1-34) over the parent strain 33C. Figure 4 summarizes sugar consumption and ethanol yield. Both 13-1-17 and 15-1-34 showed a dramatic increase in arabinose utilization compared with 33C, verifying the superior performance of the arabinose strains. For example, strain 13-1-17 was able to consume 86% and 73% arabinose while the parent strain, 33C only 64% and 37% in slurries A and B, respectively. Fermentation profiles of 13-1-17 in slurries A and B are shown in Figure 5. Co-utilization of sugars is observed though a

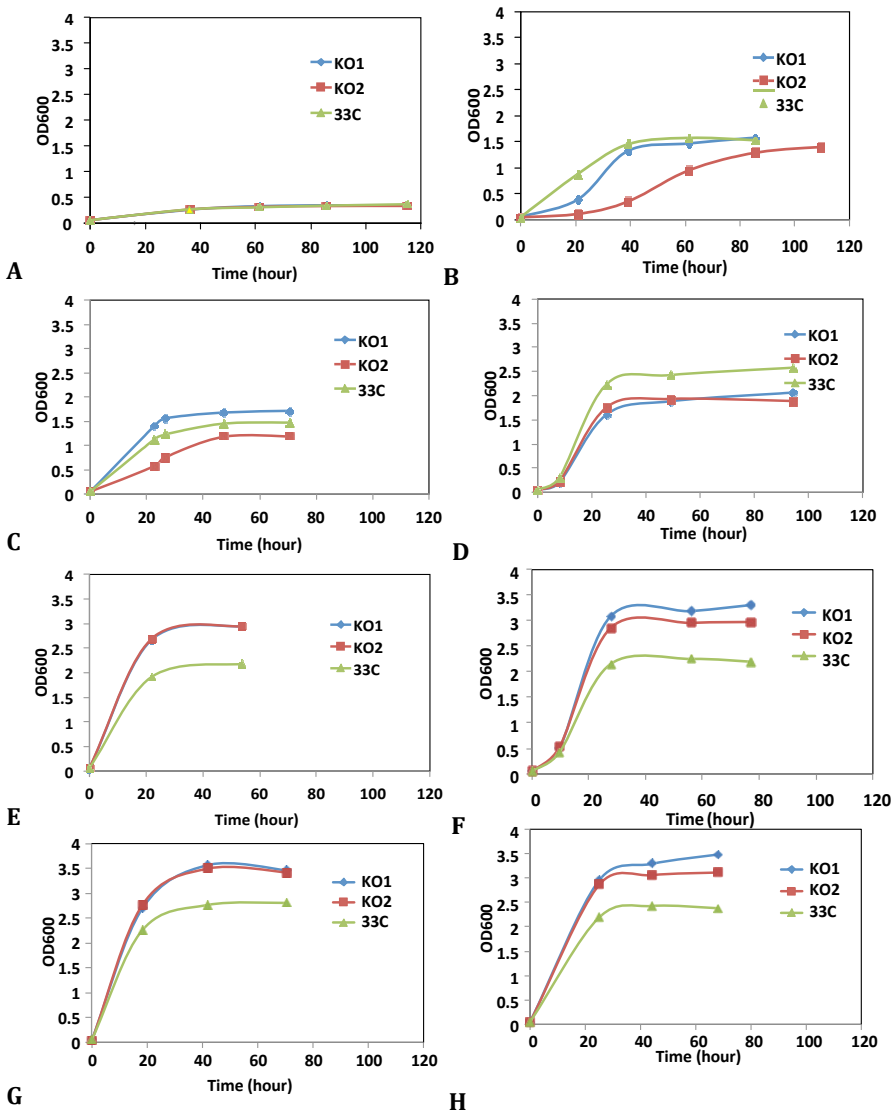

Figure 2: Serial transfers of 33C KO Library (knockout library) grown in RMA5 (35°C) shake flasks to enrich for faster growers. KO1 and KO2 were cultures revived from two vials of the frozen 33C KO library. 33C was included as a control in this enrichment growth. A, first transfer, B, third transfer, C, fifth transfer, D, ninth transfer, E, twelfth transfer, F, thirteenth transfer, G, fifteenth transfer and H, seventeenth transfer.

preference of the sugars in the order of glucose, xylose and arabinose also appears to be present, indicated by the rate of consumption. It is recognized that the ethanol titers were similar between 13-1-17 (and 15-1-34) and 33C despite increased arabinose consumption. We are currently unsure of the metabolic fate of the arabinose, but ultimately for the production of cellulosic ethanol it will be important to drive this flux towards ethanol production. We speculate that perhaps this phenomenon may be a result of the low energetic state of the cultures at the end of the fermentation, in which relatively high concentration of ethanol was already present from the conversion of glucose and xylose which are consumed more rapidly than arabinose (Figure 5). However, future flux experiments will be required to ascertain the fate of the arabinose metabolized during late-fermentation.

Genetic characterization 13-1-17 and 33C through Next Generation Sequencing

In addition to generating a superior arabinose utilizing strain, it would be very informative if we could understand the causative underlying genetic alterations leading to this phenotypic enhancement. The results would help identify the genes or regions in the genome, which are disrupted by the insertions, giving us insights into the genetic basis for improved growth. To verify the genomic insertion loci of transposon and identify other potential genetic changes besides transposon insertion occurring during arabinose selection/enrichment process, we further employed Next Generation Sequencing (NGS) technology to resequence the entire genomes of top performing arabinose-utilizing strains.

Through NGS-based genome resequencing, we located the integration loci of the transposons in the genomes of 33C and 13-1-17. Transposon *loxP-aadA-loxP-*P_{gap}*-araBAD* was inserted in a native plasmid of 33C, disrupting a hypothetical protein gene. The second transposon containing only *aadA* (created during the KO library construction) was inserted within ZMO1650 in the chromosome of 13-1-17. This latter insertion site was surprising in that we cannot envision

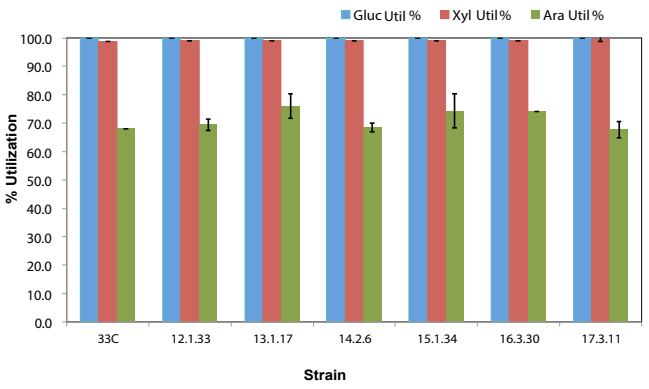

Figure 3: Mixed sugar fermentation (RMG1X4A1) of single isolates from KO library in shake flasks.

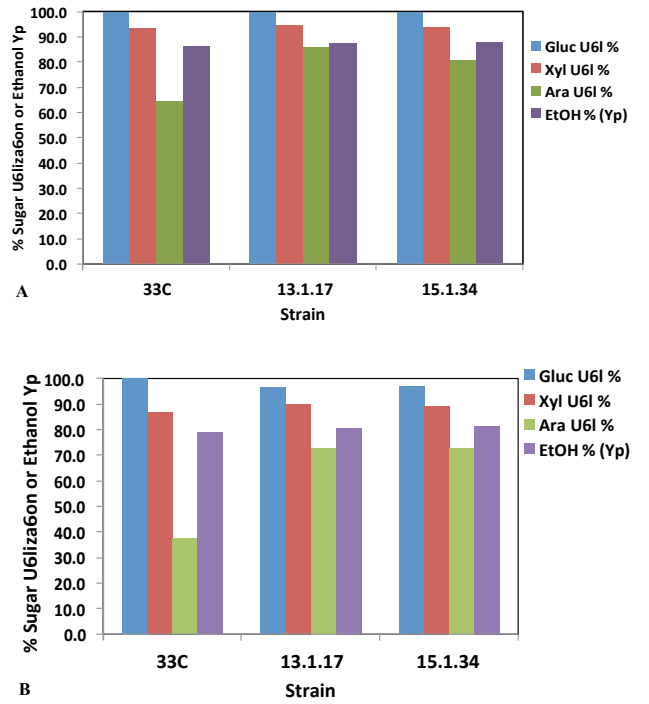

Figure 4. Sugar consumption and ethanol yield of arabinose strains in the 20%TS PCS de-acetylated, saccharified slurry A (A) and 15% TS PCS saccharified slurry B (B). Data was taken after 97 hours (for slurry A) or 125 hours (for slurry B) of fermentation. Yp: ethanol yield calculated based on total glucose, xylose and arabinose.

a scenario where the disruption of this gene, encoding a beta-lactamase would lead to improved arabinose utilization. However, NGS also showed that additional mutations had occurred in these strains whose role in enhanced arabinose utilization can be rationally understood. Accordingly, we believe that transposon insertion events have little to do with the enhanced phenotype, and rather the SNPs that occurred by chance are primarily responsible for the phenotype of enhanced arabinose utilization in this work.

SNP_s in strains 33C and 13-1-17 relative to their parents were revealed (Table 2). Specifically, mutations, which we speculated to be relevant to arabinose utilization, were identified within the *araB* itself in strains 33C and 13-1-17, suggesting the possibility that direct arabinose pathway modulation had occurred. When we compared the sequence in 33C to the published sequence of *E. coli*, [16] from which the *araBAD* was used to construct the transposon *loxP-aadA-loxP-*P_{gap}*-araBAD*, we observed that 33C had acquired 3 independent mutations in the araB gene (R127E, S128A, A490G), encoding ribulokinase. Furthermore, in 13-1-17 the A490G mutation was further altered to A490V (a G to V amino acid mutation from 33C to 13-1-17). This strongly implicates altered ribulokinase activity (either through altered protein levels or kinetic properties) as being central to the enhanced phenotype from the initial integrants (which did not grow on arabinose readily), to 33C then to 13-1-17. Additionally, a mutation in the putative Ribosome Binding Site (RBS) of the gene *rpiB* (ZMO1200, encoding ribose-5-P-isomerase, RpiB) was identified in strain 13-1-17. RpiB promotes the

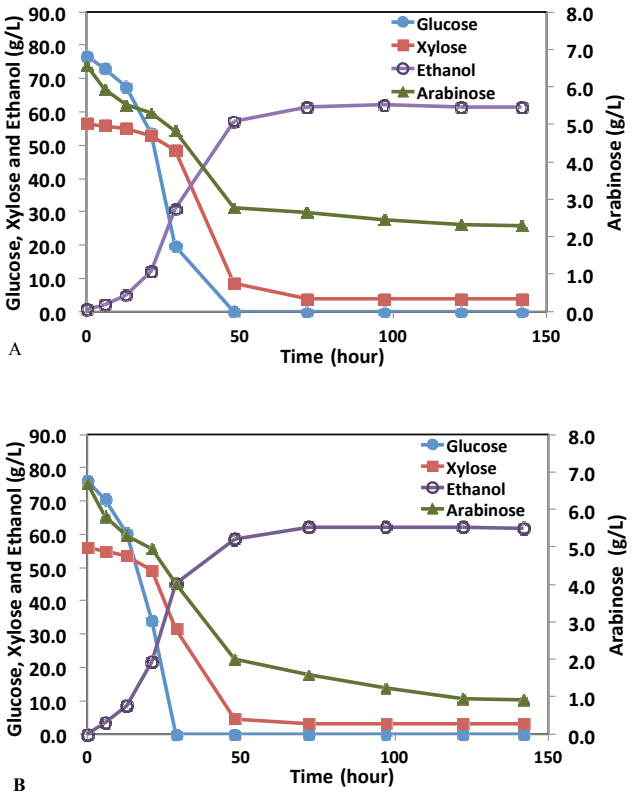

Figure 5: Fermentation profiles of 13-1-17 and parent strain 33C in 20%TS PCS saccharified slurries A and B.
(A), 33C in slurry A; (B), 13-1-17 in slurry A; (C), 33C in slurry B and (D), 13-1-17 in slurry B.

	8b	33C	13-1-17
araBAD in native plasmid (ref. 8b)	no	yes	yes
aadA in ZMO1650 (ref. 8b)	no	no	yes
araB mutation (ref. *E .coli*)	no	Yes: R127E, S128A, **A490G**	Yes: R127E, S128A, **A490V**
ZMO0353 mutation (ref. 8b)	no	no	Yes: A376V
ZMO1200 mutation (ref. 8b)	no	no	yes

ref: referenced genome sequence
ZMO1650: β--lactamase gene
ZMO0353: 4--diphosphocytidyl--2C--methyl--D--erythritol synthase gene
ZMO1200: ribose--5--phosphate isomerase Gene (*rpiB*)
Table 2: Transposon insertions and mutations identified through Next Generation Sequencing.

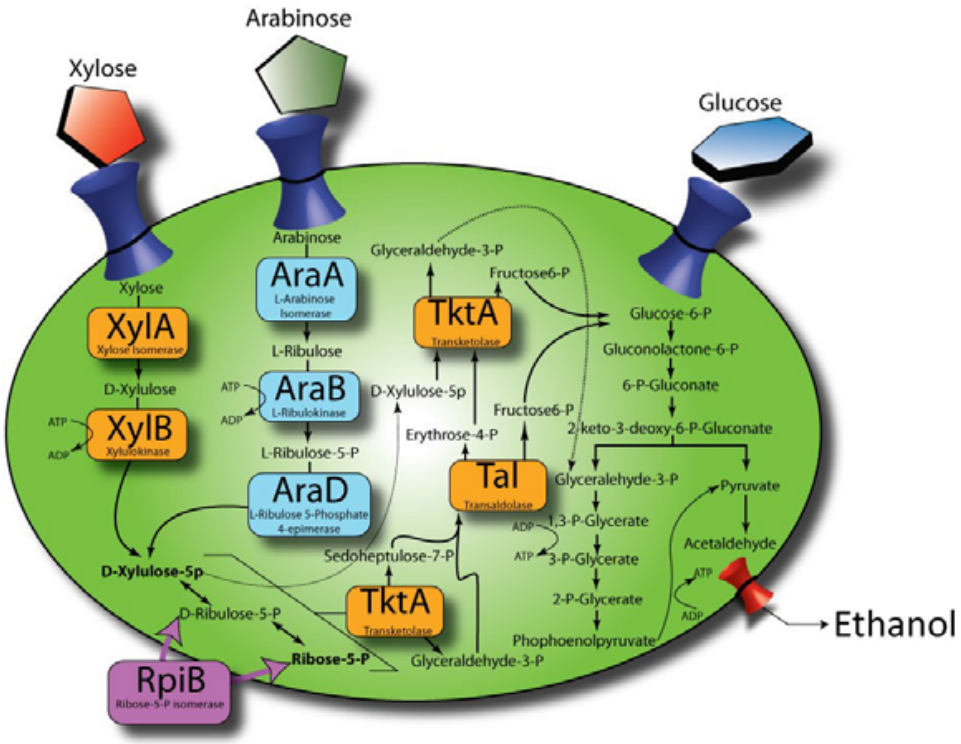

Figure 6: Pentose phospshate pathway in recombinant *Zymomonas mobilis*.

isomerization from D-ribulose-5-P to Ribose-5-P and lies right at the junction of the pentose phosphate pathway (Figure 6).

One final SNP that was identified in 13-1-17 was within ZMO0353, encoding a 4-diphosphocytidyl-2C-methyl- D-erythritol synthase (A376V), acting as an intermediate step within the methylerythritol phosphate pathway.

Discussion

We set out to utilize numerous techniques to improve arabinose utilization in our current highest performing ethanologen, *Z. mobilis* strain 8b. In order to generate this strain we attempted strategies including independent and combinatorial overexpression of the individual arabinose pathway genes, Global Transcription Machinery Engineering (GTME) [17], gene overexpression libraries (data not shown), and as described here transposon6 based genomic knockout libraries. The advantage of using transposons for mutagenesis is that a large and diverse collection of mutations can be generated in random locations throughout the genome, rendering greater chances of beneficial effect to the organisms from the mutations. Ironically, while this method yielded the highest performing strain as it relates to arabinose usage, we believe that this phenotype was not derived from the transposon integration event, but rather unrelated spontaneous mutations that occurred within the genome of this strain, or the synergetic effect of transposon integration and spontaneous mutations. However, this is speculative and has not yet been confirmed experimentally. Specifically, we identified mutations affecting three independent proteins via NGS-based genome resequencing. The relationship between the gene mutations and the desired phenotype (e.g. improved arabinose utilization) can then be tested by re-creating the insertion mutation(s) in the parental strain (e.g. 33C) and then examining the phenotype.

The first protein that was found altered in strain 13-1-17 compared to 33C was 4-diphosphocytidyl-2C-methyl-D-erythritol synthase (A376V) encoded by ZMO0353. This protein plays a key step in the methylerythritol phosphate pathway (MEP), but it is difficult to theoretically tie this mutation with the observed phenotype. However, a common metabolite link between the MEP pathway and the pentose phosphate pathway is D-glyeraldehyde-3-phosphate. It is possible that this mutation is related to metabolic flux alterations between these two pathways. However, recapitulation of this mutation, and further analysis, including flux balance analysis, will be required to understand its effect on arabinose usage.

The second subset of these mutations was located within the araB gene encoding ribulokinase that we introduced from *E. coli* leading to these translational changes: R127E, S128A, and A490G in strain 33C. While 33C is greatly inferior to its derivative strain 13-1-17 for arabinose utilization, it is markedly improved over the very first *araBAD* transformant/integrant, selected on Sp (Supplementary Figure S1). This integrant was cultured in arabinose containing medium, which likely provided the selective pressure on this strain to isolate the mutations found within 33C. Interestingly, compared to 33C, strain 13-1-17 has an additional mutation to araB, which again targeted amino acid residue 490 (G490V). It will be interesting to assay these various mutant forms of araB to understand their effect on the ribulokinase activity, and pathway flux as a whole.

The third mutation we identified in 13-1-17 (compared to 33C) was located within the predicted RBS of the ribose 5-P-isomerase (RpiB) encoded by ZMO1200. As this gene functions within the pathway of pentose conversion to ethanol, we reasoned that this gene influenced flux through the pathway. RpiB plays a role in the isomerization of ribulose-5-P to ribose 5-P. We hypothesize that insufficient levels of

ribose-5-P represent a bottleneck within this pathway in strain 33C, which was then upregulated by the *rpiB* mutation in strain 13-1-17. This is supported by work reported in Caimi et al. [18], which suggests that *rpiB* overexpression leads to improved xylose usage. Given that arabinose usage and xylose usage converge at a branch-point further up in the pathway than where RpiB acts (Figure 6), it is logical that rpiB overexpression would also lead to improved arabinose usage. To ascertain whether this mutation leads to an increased or decreased expression of *rpiB*, a detailed activity assay to compare strains13-1-17 and its parent, 33C, for this enzyme will be called for.

Acetate has been shown to be inhibitory to the growth and fermentation of *Z. mobilis* [15]. Our results confirmed this observation in that all strains performed worse in the non de-acetylated hydrolyzate (Figure 4B) compared with in the de-acetylated hydrolyzate (Figure 4A). It is noteworthy that the improvement on arabinose utilization yield was greater for strains 13-1-17 and 15-1-34 compared with 33C when tested in the non de9 acetylated hydrolyzate. Though there is no identified mutation(s) that can explain the acetate toxicity in these strains, we suspect that acetate played a role in the performance difference between the improved and the parental strains. Further investigation in this area (e.g. examining the by-product formation profile) will be important to understand the relationship between arabinose utilization efficiency and acetate toxicity.

Regardless of the genetic mechanisms underlying the phenotypic enhancements of these strains, the functional outcome is that through the methods we employed, we have generated a strain of *Z. mobilis* that is excellent in its ability to utilize glucose, xylose and arabinose, while simultaneously producing high titers and yields of ethanol. Furthermore, these traits were shown to be relevant when using an industrial relevant feedstock, saccharified slurry of dilute acid pretreated corn stover. The ability of strain 13-1-17 to utilize 100% of the glucose, 95% of the xylose and over 85% of the arabinose in a 20% TS hydrolyzate represents a significant step forward in the realization of economically viable cellulosic ethanol.

Conclusions

A glucose, xylose and arabinose utilizing *Zymomonas mobilis* strain was constructed by incorporating arabinose catabolic pathway genes, *araBAD* in a glucose, xylose co-fermenting host, 8b, using a transposition integration approach. Through a second transposition to generate a mutagenic knockout library and the selection/enrichment culturing process, strain 13-1-17 was isolated. Subsequent fermentation evaluation indicated significant improvement in metabolizing arabinose in dilute acid pretreated, saccharified corn stover slurry. Next Generation Sequencing of the genomic DNA from strain 13-1-17 suggested several mutations, which may have impacted the enhanced fermentation performance.

Acknowledgments

This work was funded by Bioenergy Technologies Office of U.S. Department of Energy. The authors would like to thank Ali Mohagheghi, Andrew Lowell, and Robert Nelson for preparing the hydrolyzates, and Philip T. Pienkos for critical reading of this manuscript.

References

1. Lawford HG (1988) A new approach to improving the performance of Zymomonas in continuous ethanol fermentations. Appl Biochem Biotechnol 17: 203-219.
2. Zhang M, Chou YC, Howe W, Eddy C, Evans K, et al. (2007) Zymomonas pentose-sugarfermenting strains and uses thereof. Patent 7: 223-575.
3. Mohagheghi A, Dowe N, Schell D, Chou YC, Eddy C, et al. (2004) Performance of a newly developed integrant of Zymomonas mobilis for ethanol production on corn stover hydrolysate. Biotechnol Lett 26: 321-325.
4. Humbird D, Davis R, Tao L, Kinchin C, Hsu D, et al. (2011) Process Design and Economics for Biochemical Conversion of Lignocellulosic Biomass to Ethanol: Dilute-Acid Pretreatment and Enzymatic Hydrolysis of Corn Stover. NREL/TP-5100-47764. Golden, CO: National Renewable Energy Laboratory. Deanda K, Zhang M, Eddy C, Picataggio S (1996) Development of an arabinose-fermenting Zymomonas mobilis strain by metabolic pathway engineering. Appl Environ Microbiol 62: 4465-4470.
5. Deanda K, Zhang M, Eddy C, Piccataggio S (1996) Development of an arabinose-fermenting Zymomonas mobilis strain by metabolic pathway engineering. Appl Environ Microbiol 62: 4465-4470.
6. Lawford HG, Rousseau JD, Tolan JS (2001) Comparative ethanol productivities of different Zymomonas recombinants fermenting oat hull hydrolyzate. Applied Biochemistry and Biotechnology 91-93: 133-146.
7. Yang JJ (2011) Zymomonas with improved arabinose utilization. US20110143408 A1.
8. Dubois V, Parizano MP, Arpin C, Coulange L, Bezian MC, et al. (2007) High genetic stability of integrons in clinical isolates of Shigella spp. of worldwide origin. Antimicrob Agents Chemother 51: 1333-1340.
9. Hoffman LM, Jendrisak JJ, Meis RJ, Goryshin IY, Reznikof SW (2000) Transposome insertional mutagenesis and direct sequencing of microbial genomes. Genetica 108: 19-24.
10. Sternberg N, Hamilton D (1981) Bacteriophage P1 site-specific recombination. I. Recombination between loxP sites. J Mol Biol 150: 467-486.
11. Yang S, Land ML, Klingeman DM, Pelletier DA, Lu TY, et al. (2010) Paradigm for industrial strain improvement identifies sodium acetate tolerance loci in Zymomonas mobilis and Saccharomyces cerevisiae. Proceedings of the National Academy of Sciences of the United States of America. 107: 10395-10400.
12. Yang S, Pappas KM, Hauser LJ, Land ML, Chen GL, et al. (2009) Improved genome annotation for Zymomonas mobilis. Nat Biotechnol 27: 893-894.
13. Agrawal M, Mao Z, Chen RR (2011) Adaptation yields a highly efficient xylose-fermenting Zymomonas mobilis strain. Biotechnol Bioeng 108: 777-785.
14. Meijnen JP, de Winde JH, Ruijssenaars HJ (2008) Engineering Pseudomonas putida S12 for efficient utilization of D-xylose and L-arabinose. Appl Environ Microbiol 74: 5031-5037.
15. Franden MA, Pilath HM, Mohagheghi A, Pienkos PT, Zhang M (2013) Inhibition of growth of Zymomonas mobilis by model compounds found in lignocellulosic hydrolysates. Biotechnol Biofuels 6: 99.
16. Lee N, Gielow W, Martin R, Hamilton E, Fowler A (1986) The organization of the *araBAD* operon of Escherichia coli. Gene 47: 231-244.
17. Alper H1, Stephanopoulos G (2007) Global transcription machinery engineering: a new approach for improving cellular phenotype. Metab Eng 9: 258-267.
18. Caimi PG, McCole L, Tao L, Tomb JF, Viitanen PV (2012) Xylose 1 utilization in recombinant Zymomonas. US 2012/0156746 A1.

Citation: Chou YC, Linger J, Yang S, Zhang M (2015) Genetic Engineering and Improvement of a Zymomonas mobilis for Arabinose Utilization and Its Performance on Pretreated Corn Stover Hydrolyzate. J Biotechnol Biomater 5: 179. doi:http://dx.doi.org/10.4172/2155-952X.1000179