

Algal Physiology: a catch all

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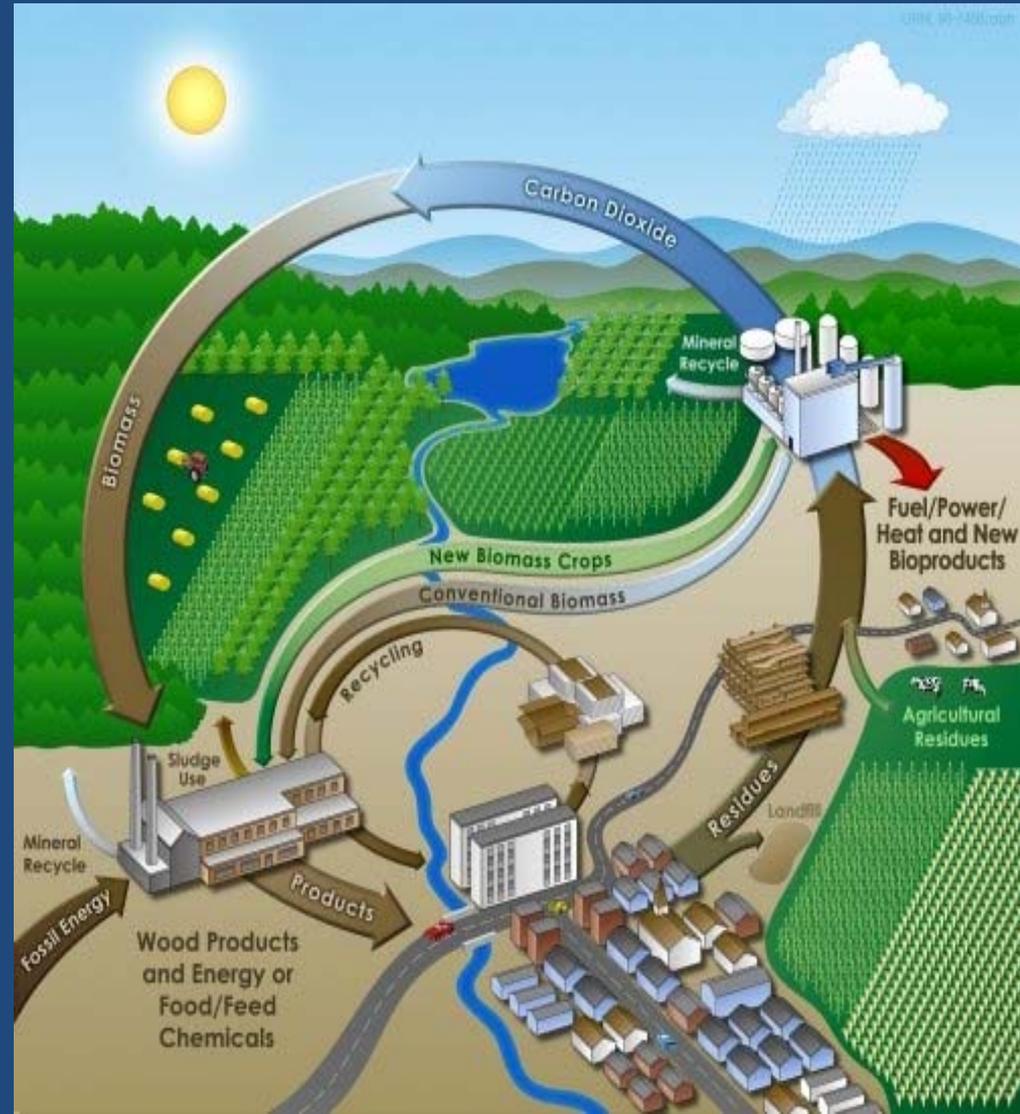
Current Projects

- Development of *Chlorella protothecoides* chloroplast and nuclear transformation vectors.
- Genetic manipulation of photosynthetic efficiency and lipid production.
- Development of phase-shifting dyes to enhance PAR.
- Development of novel lipid extraction technologies.



Outline

- Choosing the right algae; biomass and oil potential.
- Heterotrophic growth boosts oil yield.
- Metabolic strategies for ameliorating stress
- Growth inhibitors; the need for waste product removal.
- Non-destructive oil extraction from continuous cultures.

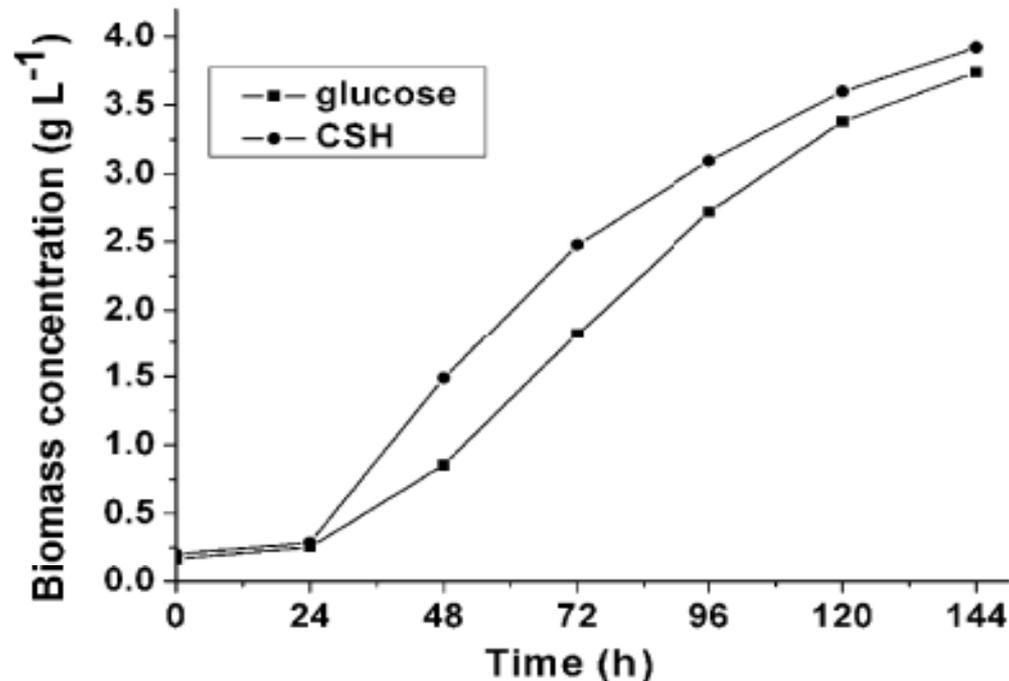


Optimizing biofuel production from microalgae

- 🕒 **Fast:** Identify the best algal strains for each locale.
- 🕒 **Fat:** Enhance lipid accumulation in microalgae having high biomass production capabilities.
- 🕒 **Facile:** Continuous (24/7), non-destructive harvesting of oils from live cultures. Development of closed-loop production systems

Why *Chlorella protothecoides*?

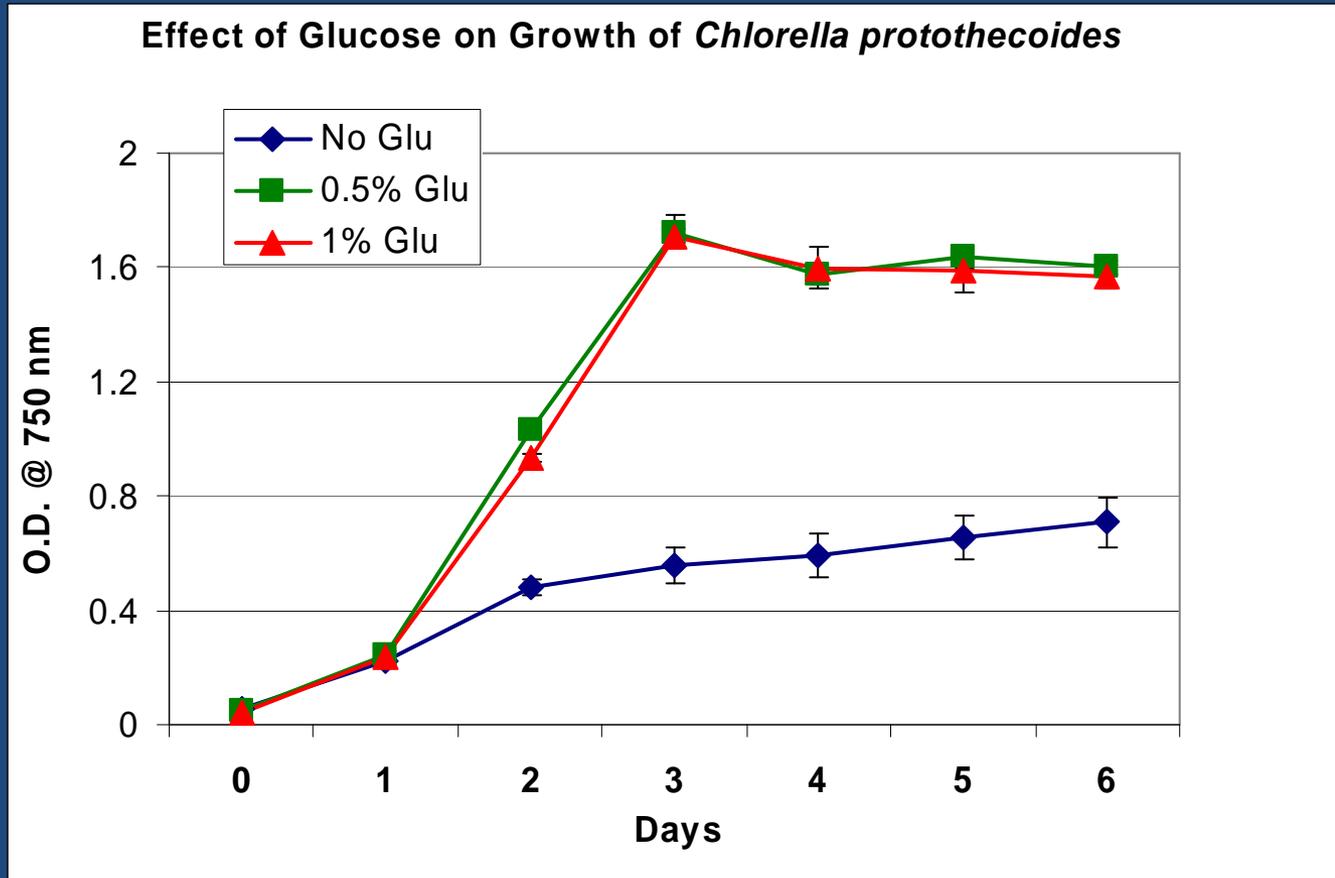
1. *C. protothecoides* has a high growth rate and high maximum culture cell density (10^8 cells/mL)
2. *C. protothecoides* is able to grow heterotrophically on glucose. Allows for growth at night (no CO_2 fixed) and greater oil production.



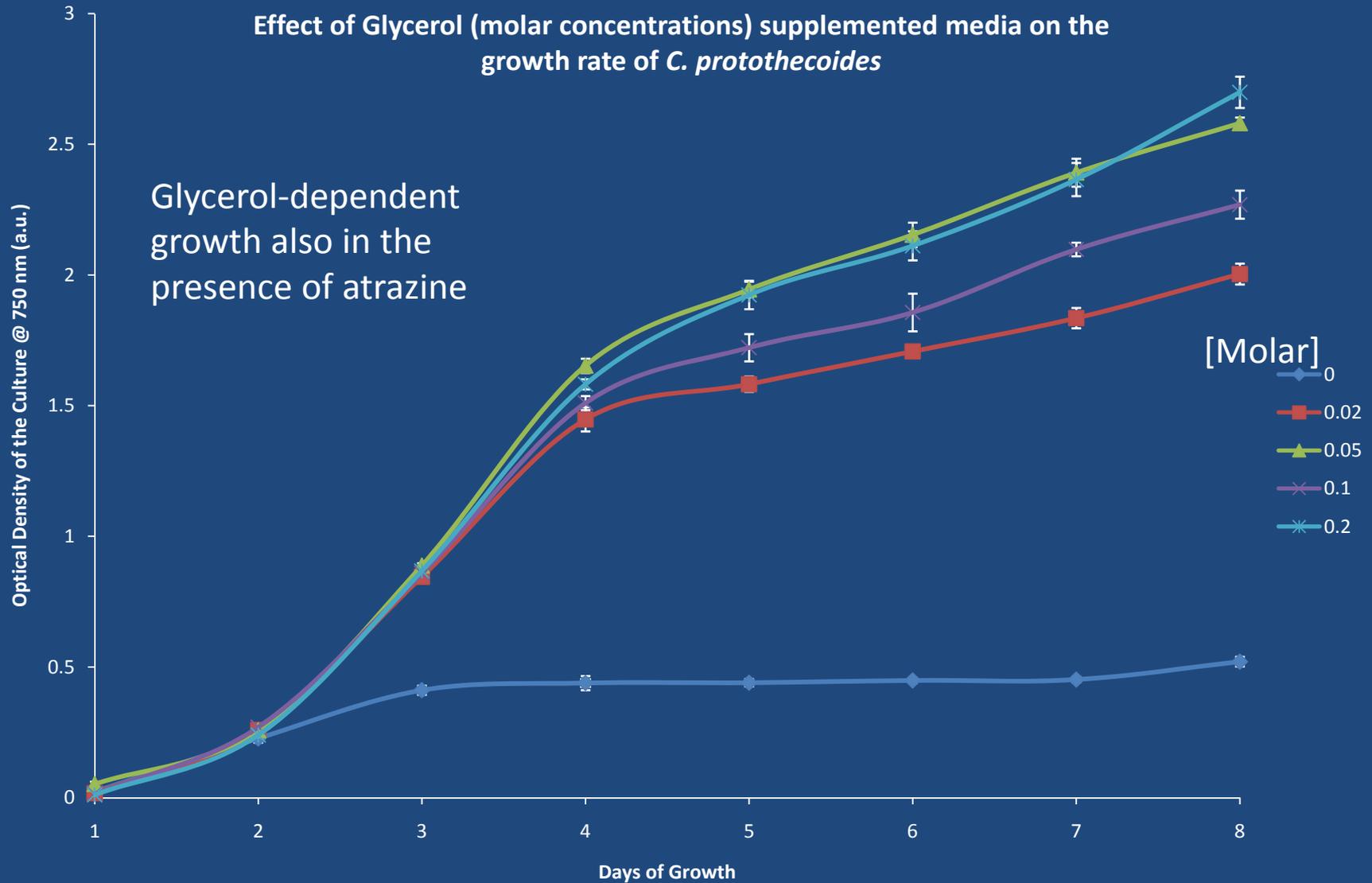
- With glucose, *C. protothecoides* has 5X the growth rate of *Chlamydomonas*.

- Biomass yields > 30 g/L have been achieved under heterotrophic conditions.

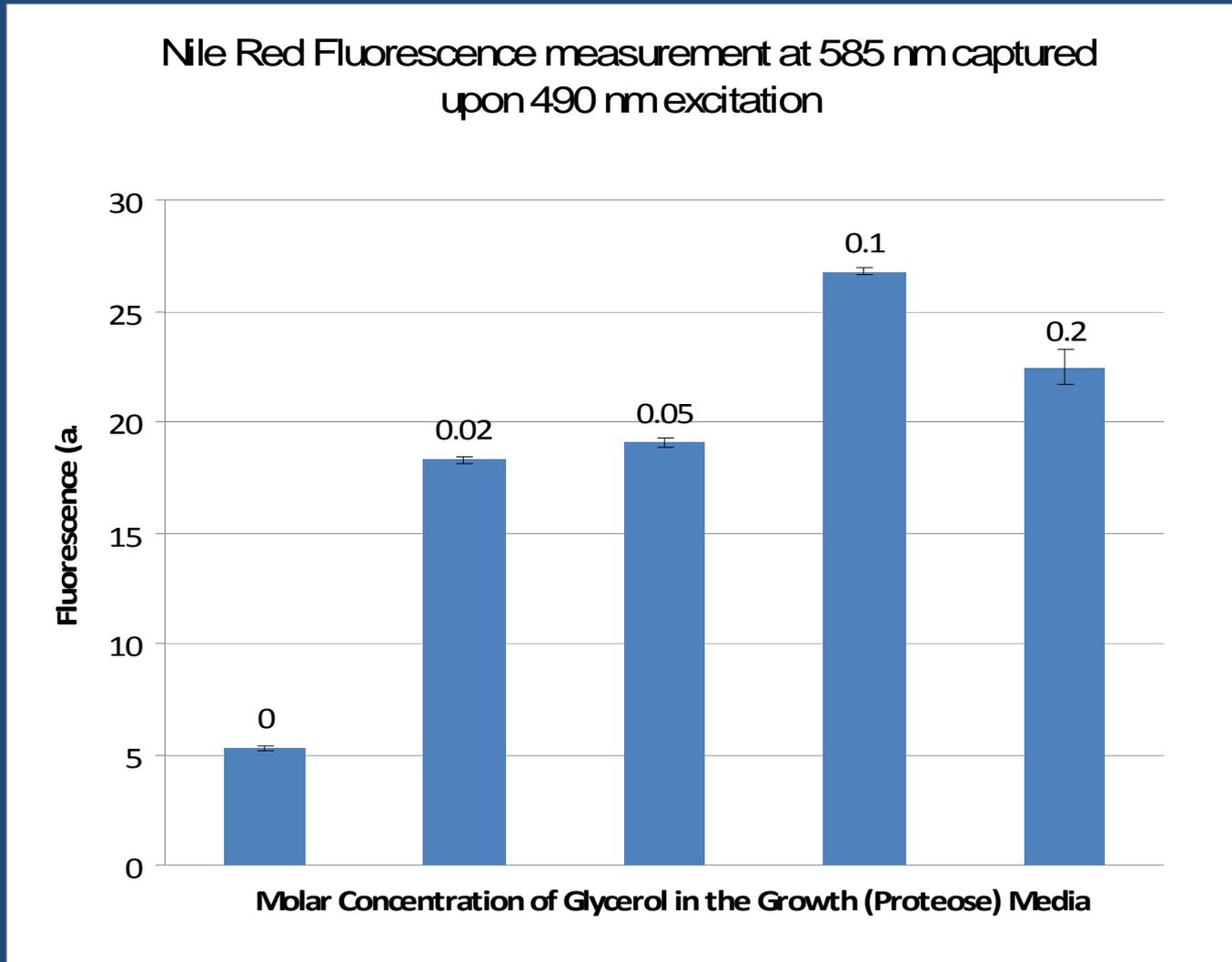
Initial doubling time is 4 times faster with glucose



C. protothecoides growth is enhanced by glycerol; a byproduct of biodiesel



Oil yield is also enhanced by glycerol



Glycerol is more efficiently converted into biomass, but glucose yields the most oil

Relative total lipid yield

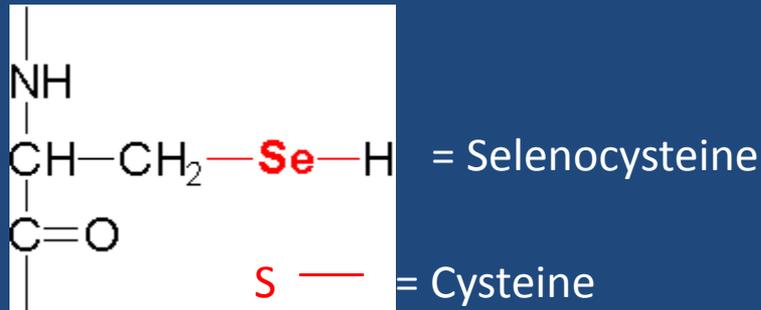
<u>Growth</u>	<u>Lipid content</u>	<u>Dry weight</u>	<u>Lipid yield</u>
No addition	5 NRU*	0.4 g/L	2 (1X)
Glycerol [20 mM]	18	1.7 g/L	30 (15X)
Glucose [15 mM]	55	1.9 g/L	105 (52X)

*NRU = Nile red units/cell

Metabolic strategies for ameliorating stress

- Selenoproteins as redox mediators, engineering selenocysteine into proteins.
- Proline, a general ROS scavenger.

Selenocysteine proteins in green algae; super redox catalysts



Selenocysteine can replace cysteine in proteins of certain organisms.

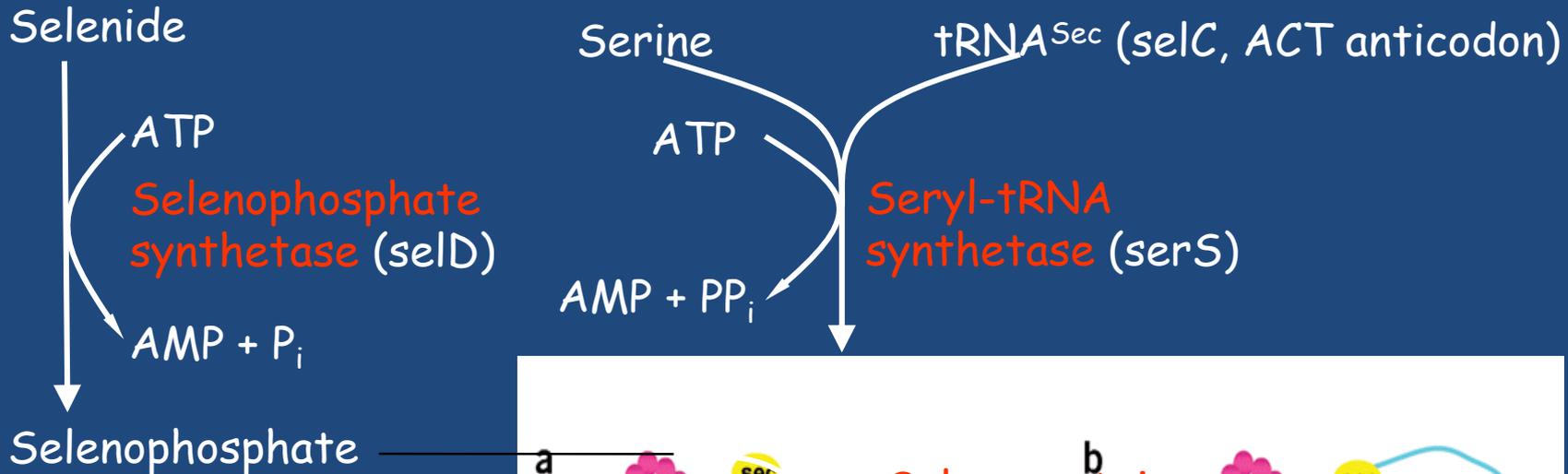
Amino Acid	pKa (pH)	E_M (mV)
Selenocysteine	5.7	- 488
Cysteine	8.3	-270

The replacement of serine or cysteine with selenocysteine in the active site of many enzymes often results in substantial increases (100-500 fold) in catalytic activity.

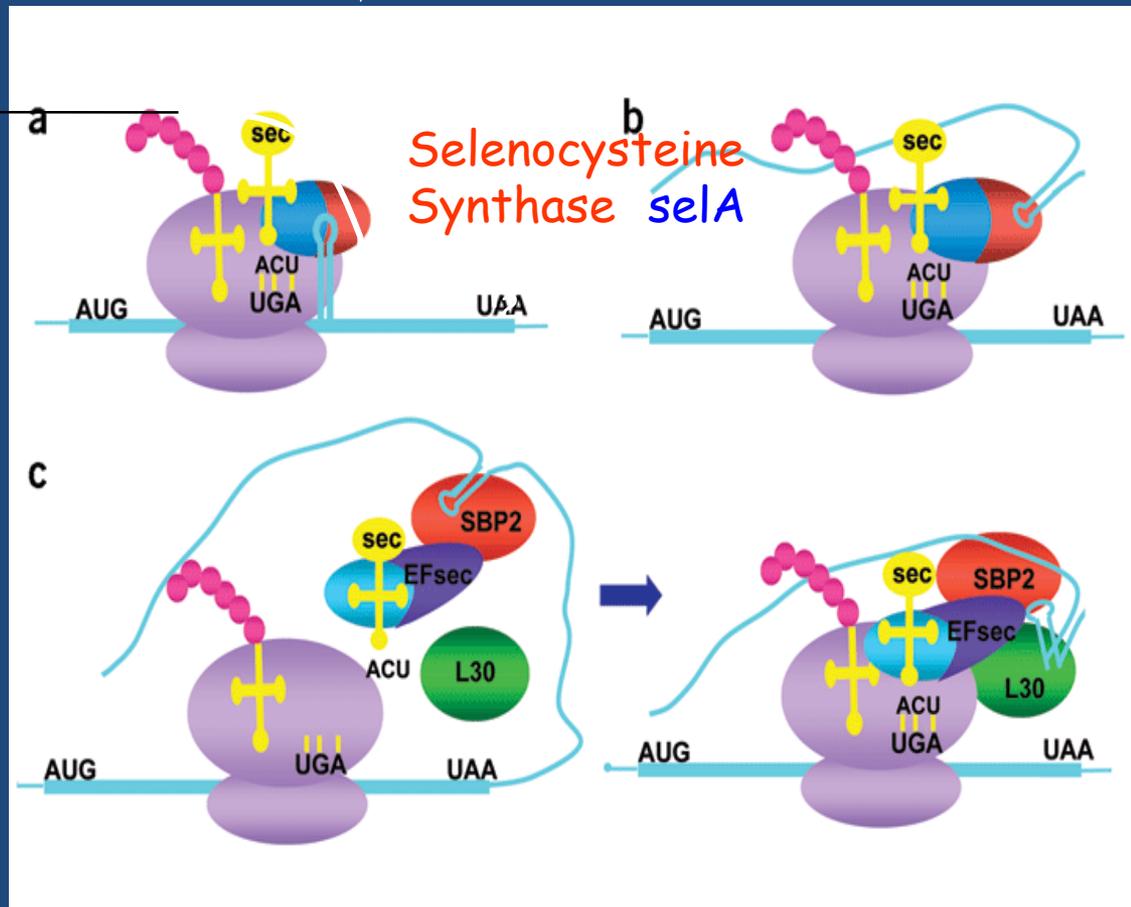
Selenocysteine has a more physiological ionization potential (pKa) and is a stronger nucleophile than cysteine.

Selenocysteine proteins in Chlamydomonas and Humans						
Gene	Function	Species	SECIS Type and length (nt)	Sec residue/ protein total length (residue)	Distance between Sec and SECIS (nt)	Stop codon used. Distance between stop codon and SECIS (nt)
<i>TR1</i>	Thioredoxin reductase	<i>C. r.</i>	I (57) (105)*	532/533	687 (528)*	UAA (681) (522)*
<i>TR1</i>	Thioredoxin reductase 1	<i>H. s.</i>	105	498/499	213	UAA (207)
<i>MsrA1</i>	Methionine-S-sulfoxide reductase	<i>C. r.</i>	II (60)	20/160	557	UGA (134)
<i>PHGPx1</i>	Phospholipid hydroperoxide Glutathione peroxidase	<i>C. r.</i>	II (107)	75/201	1055	UAA (674)
<i>PHGPx2</i>	Phospholipid hydroperoxide glutathione peroxidase	<i>C. r.</i>	II (114)	100/267	855	UAG (351)
<i>SELK1</i>	Selenoprotein K	<i>C. r.</i>	I (93)	91/92	396	UAA (390)
<i>SELK</i>	Selenoprotein K	<i>H. s.</i>	(103)	92/94	120	UAA (111)
<i>SELM1</i>	Selenoprotein M	<i>C. r.</i>	II (100)	46/140	328	UGA (43)
<i>SELM2</i>	Selenoprotein M	<i>C. r.</i>	II (64)	33/138	421	UAG (103)
<i>SELW1</i>	Selenoprotein W	<i>C. r.</i>	II (100)	14/88	406	UAA (181)
<i>SELW2</i>	Selenoprotein W	<i>C. r.</i>	II (94)	16/80	676	UGA (481)
<i>SELW</i>	Selenoprotein W	<i>H. s.</i>	(96)	13/87	256	UAA (31)
<i>GPX1</i>	Glutathione peroxidase 1	<i>H. s.</i>	(95)	49/203	505**	UAG (40)

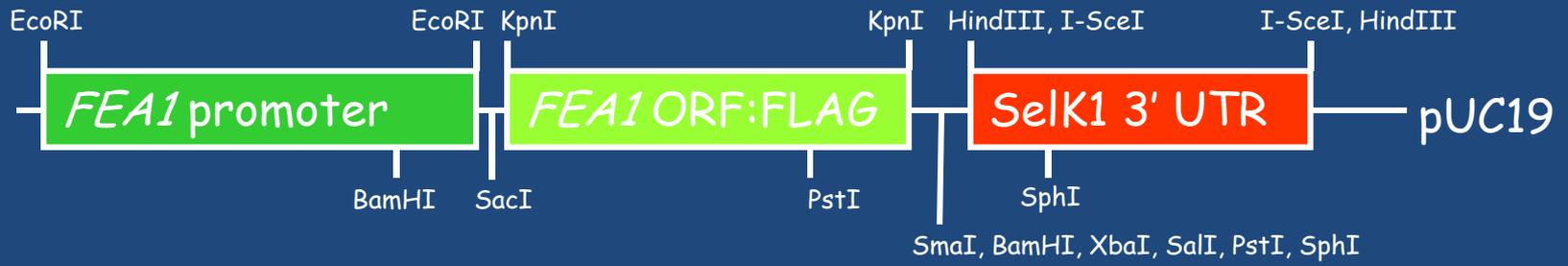
Expressing selenoproteins in *Chlamydomonas*



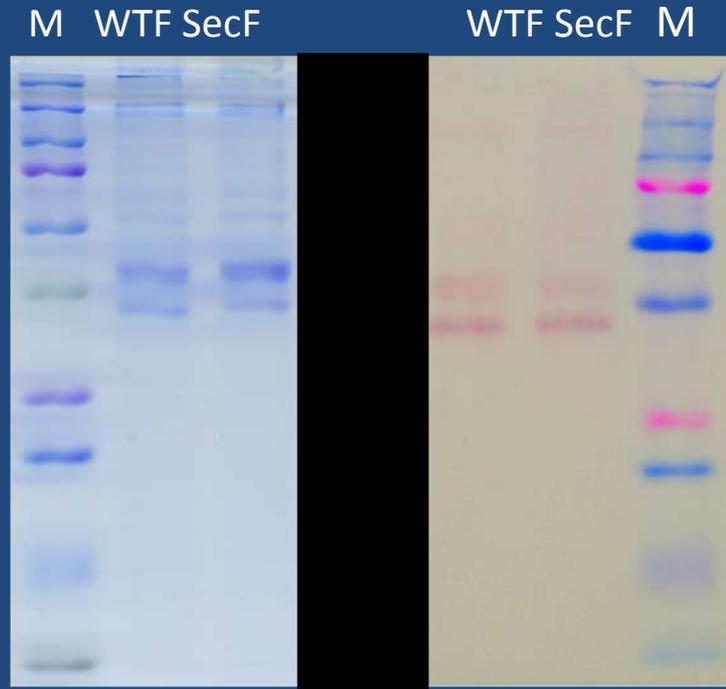
Selenophosphate



Expression of recombinant, FLAG-tagged, Fea1 selenoprotein (yield = 2 mg/L).



SDS-PAGE



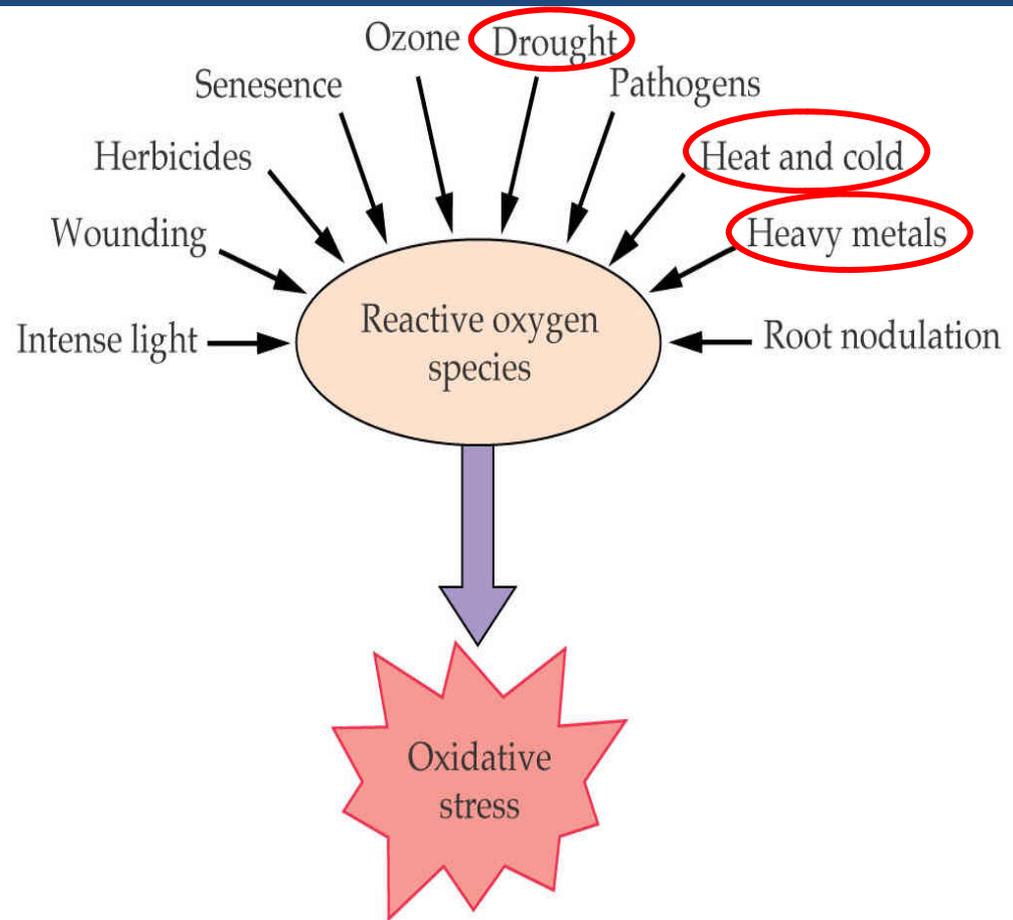
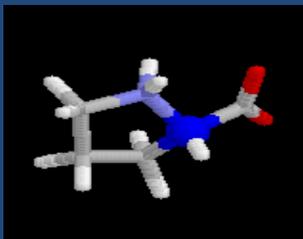
western

Proline Accumulation

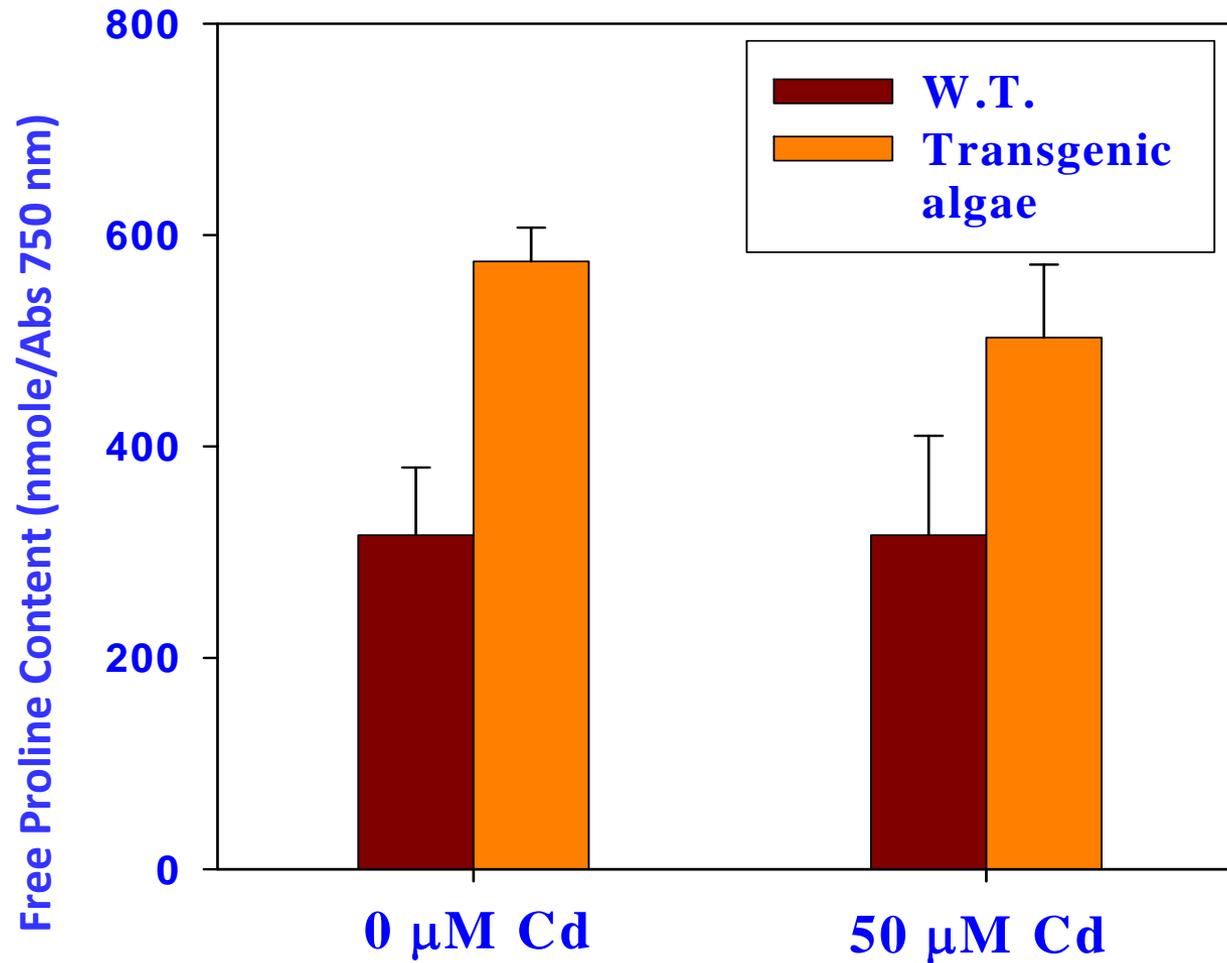
Induction of Oxidative Stress

1. Drought Stress
2. Salinity Stress
3. Cold Stress
4. Heavy Metal Stress
5. Oxidative Stress

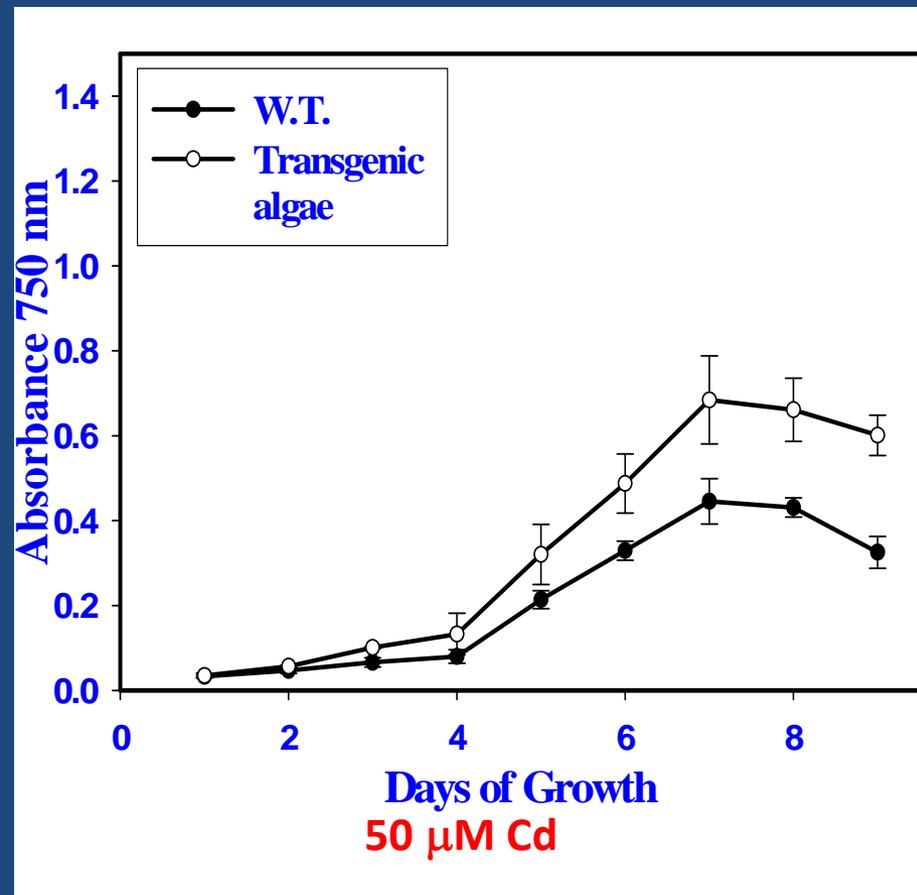
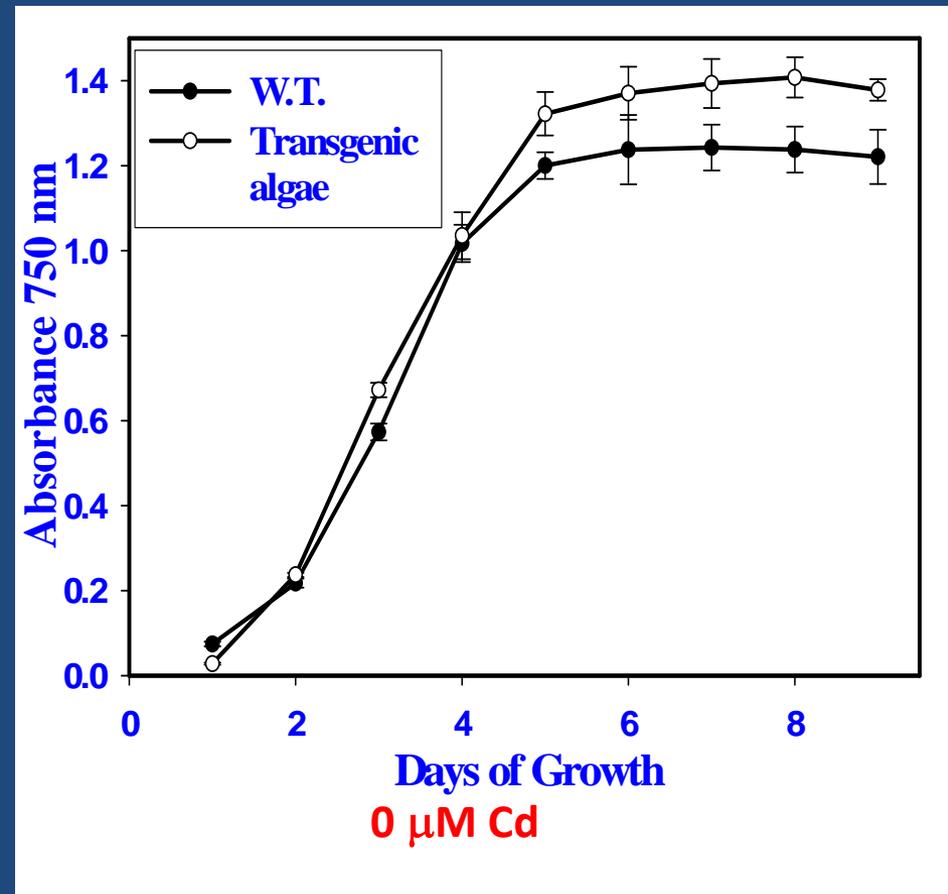
Proline



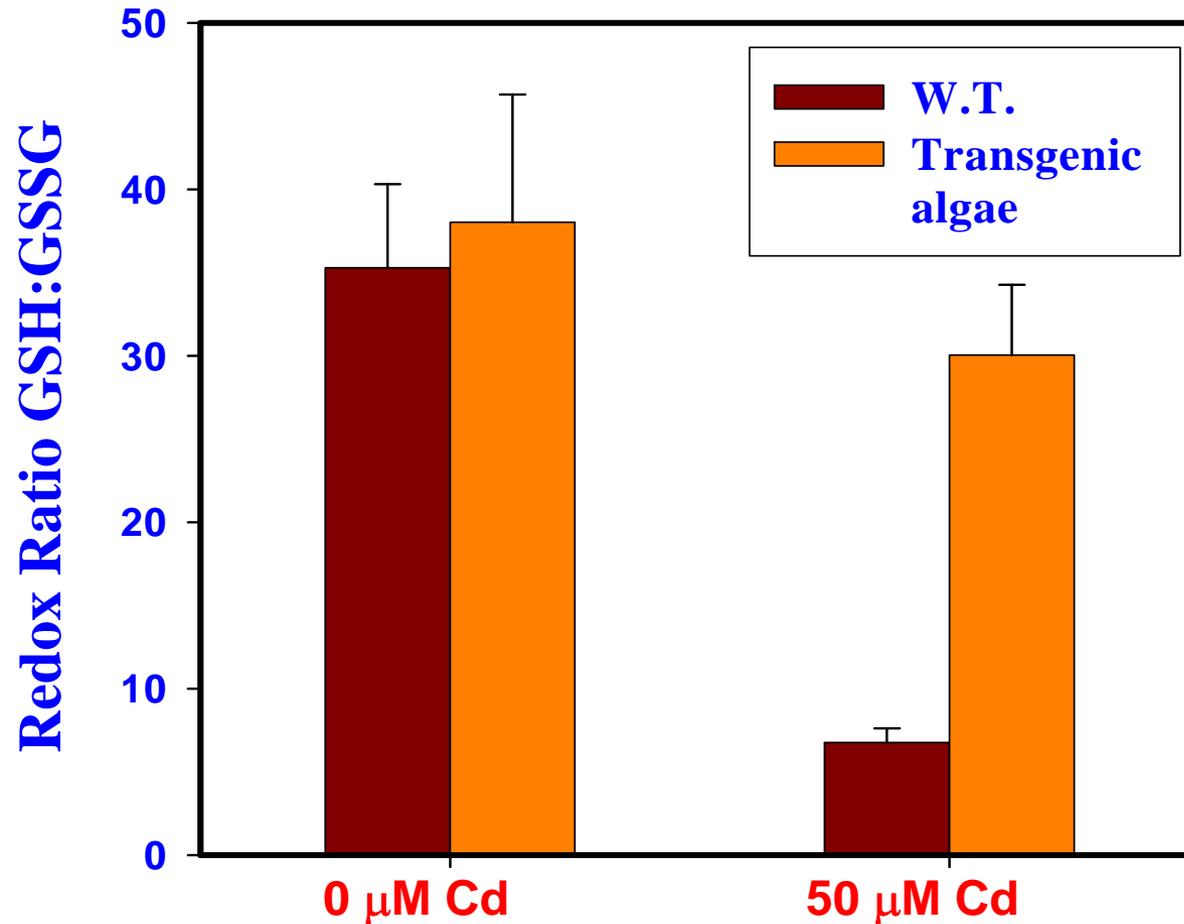
Free proline content of wild-type and transformed *Chlamydomonas* cells expressing foreign *P5CS* gene



Growth of wild-type (CC-425) and transformed *Chlamydomonas* expressing the *P5CS* gene



GSH:GSSG ratio is 4-fold higher in transgenic algae under stress



Growth inhibitors secreted by algae

- *Chlorellin*, first described from media filtrate in 1940.
- Soluble in organic solvents.
- Inactivated by 2 hr boiling.
- *Chlorellin* is a mixture of linoleic and linolenic acids.

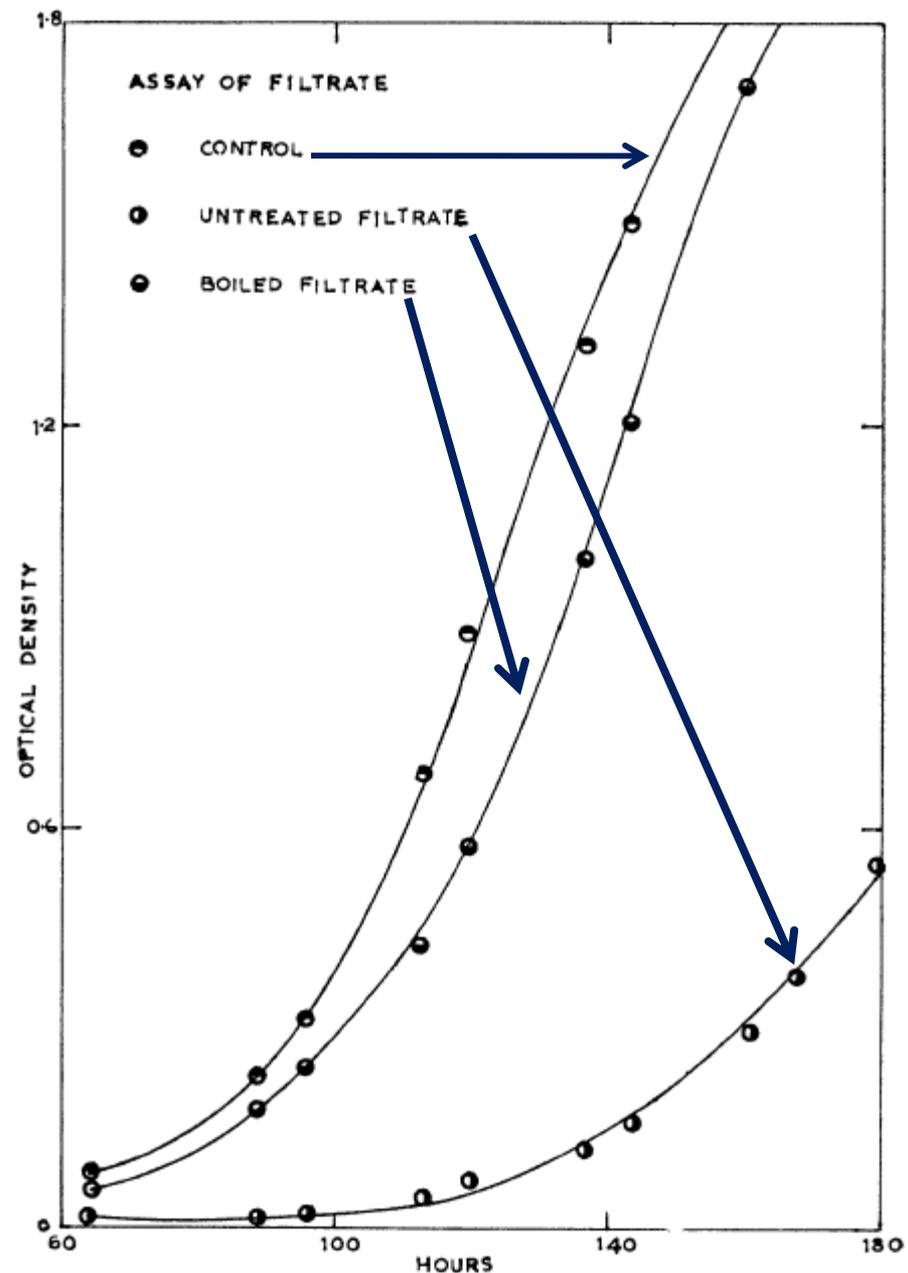


Fig. 1. Assay of filtrates of *Chlorella*.

Am. J. Bot. 27, 52 (1940)

Am. J. Bot. 51, 581 (1964)

Sensitivity of algal growth to chlorellin

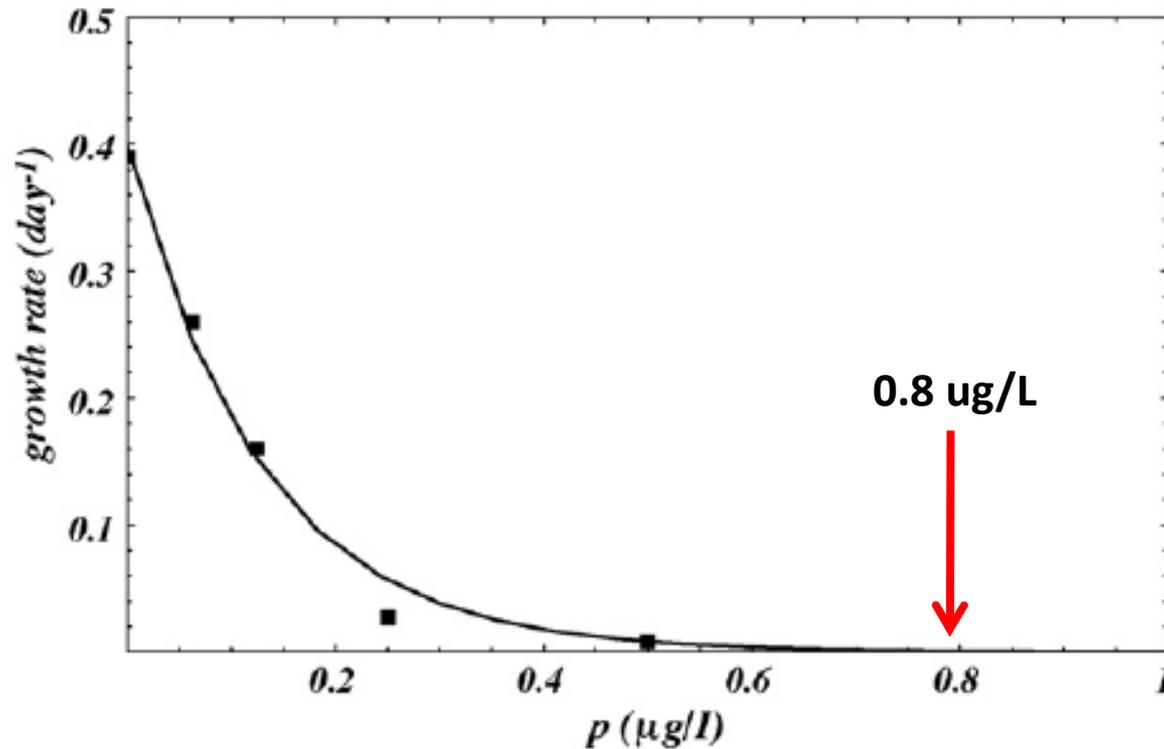


Fig. 3 – Dependence of *P. subcapitata* growth rate on the chlorellin concentration p . Comparison between experimental data and the exponential function $f_1(\bar{S}) e^{-\gamma p}$ where $\gamma = 7.81$ ($\mu\text{g/l}$).

Growth inhibition by *chlorellin* is reduced by media dilution or by activated charcoal

Algae were either grown in media (1, 2) or in dialysis bags placed in 5 volumes of external media (3, 4).

The external media (4) was either renewed (50%/day) or activated charcoal (5g/L) was added (3).

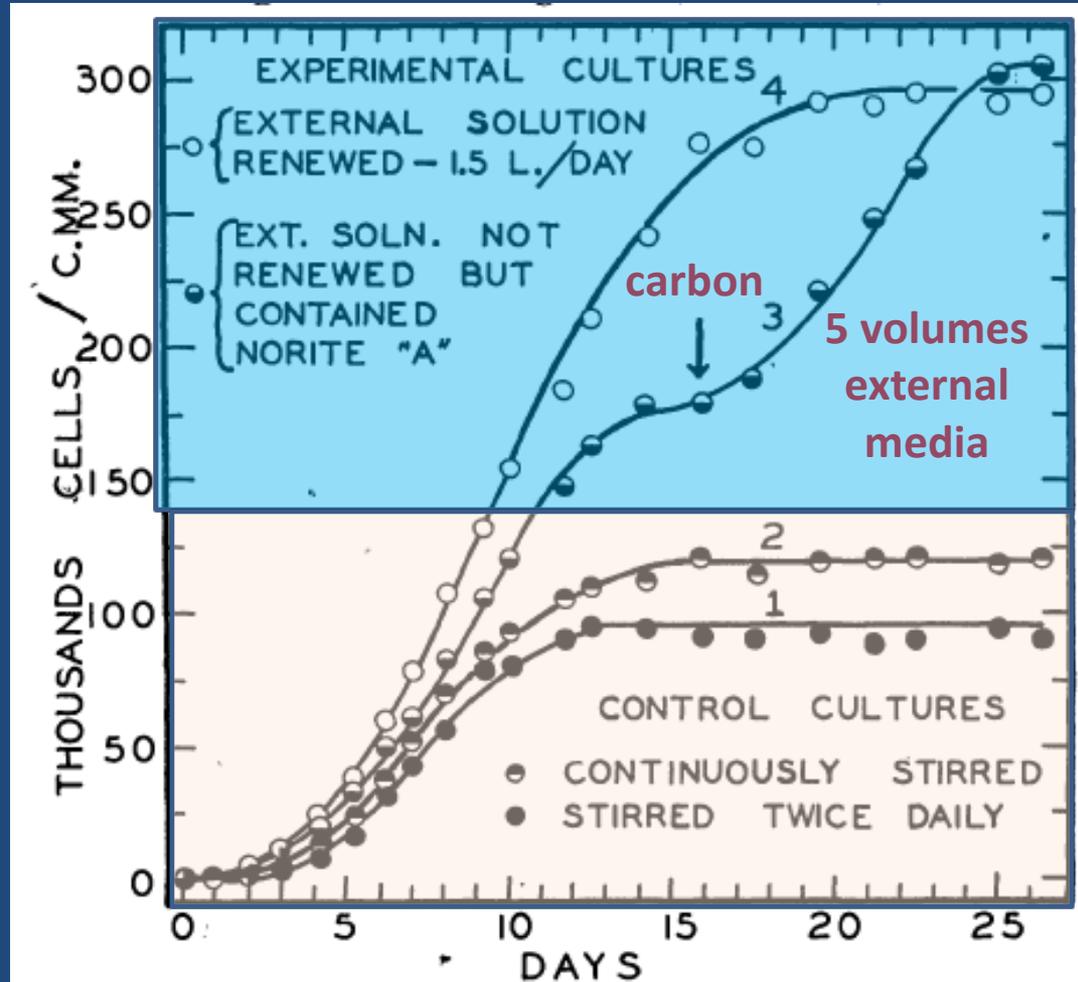


Fig. 1. Growth of *Chlorella vulgaris* in collodion sacs under different experimental conditions. Arrow in curve 3 indicates time at which Norite A was added to external solution. (See text for full explanation.)

Reducing the costs of harvesting oils from algae

Harvesting, rupturing, drying and extracting oils from algae accounts for 40-60% of the cost of producing biodiesel and places additional demands on culture replenishment.

There is a need for a low cost oil extraction technology

Non-destructive lipophile extraction: "milking" algae

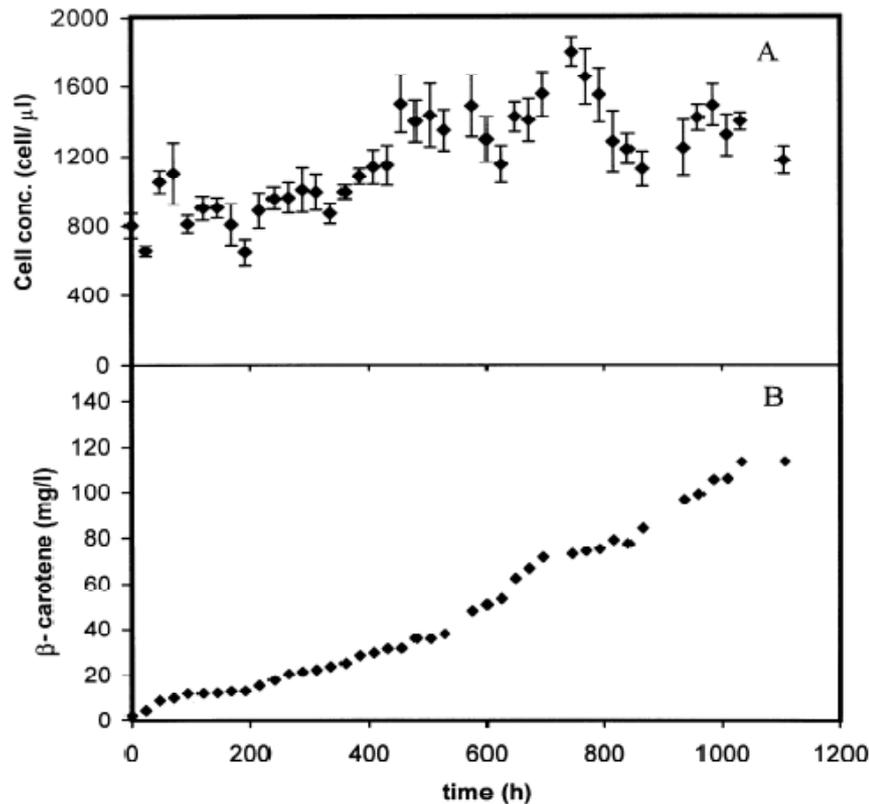


Figure 3. Growth (A) and total volumetric production of β -carotene (B) by *D. salina* in the presence of organic biocompatible solvent. Error bars show 95% confidence interval of triplicate samples taken from the bioreactor.

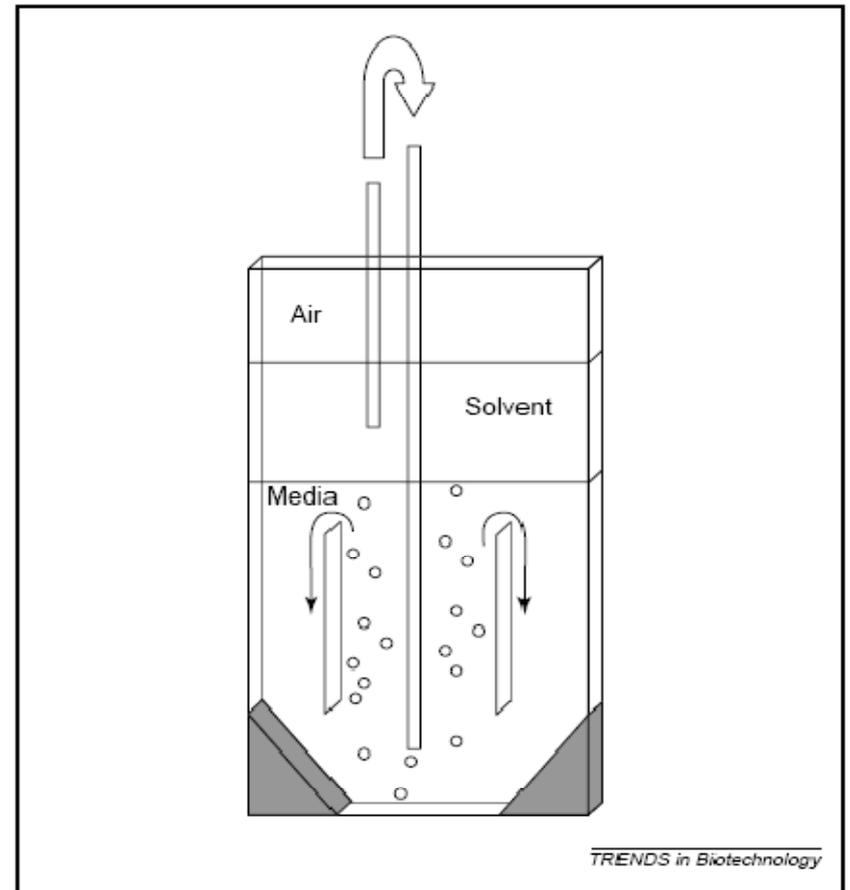
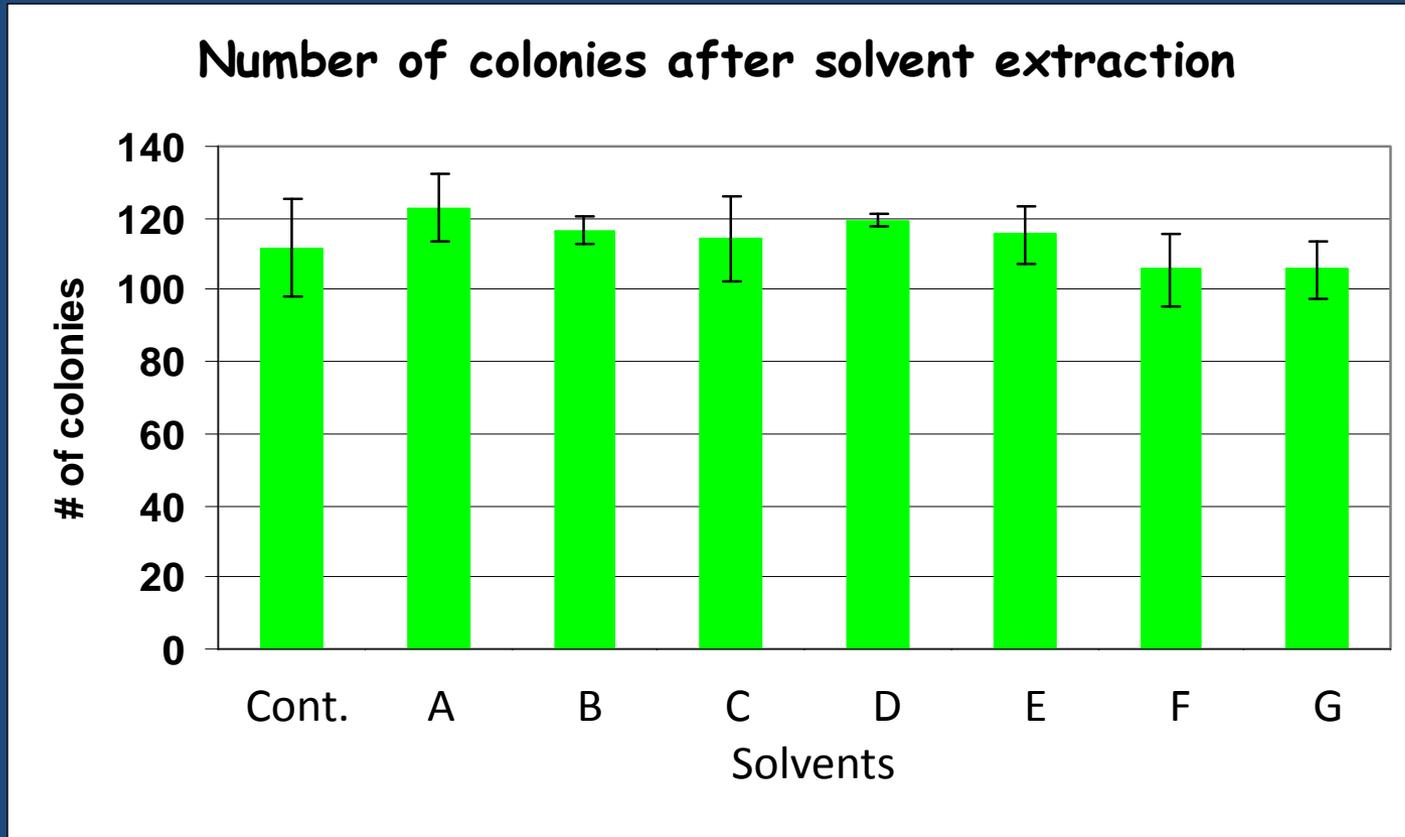


Figure 1. Schematic representation of a flat panel two-phase bioreactor used in the milking process of *Dunaliella salina* for β -carotene production. An organic phase is continuously re-circulated through the aqueous phase, resulting in extraction of the product.

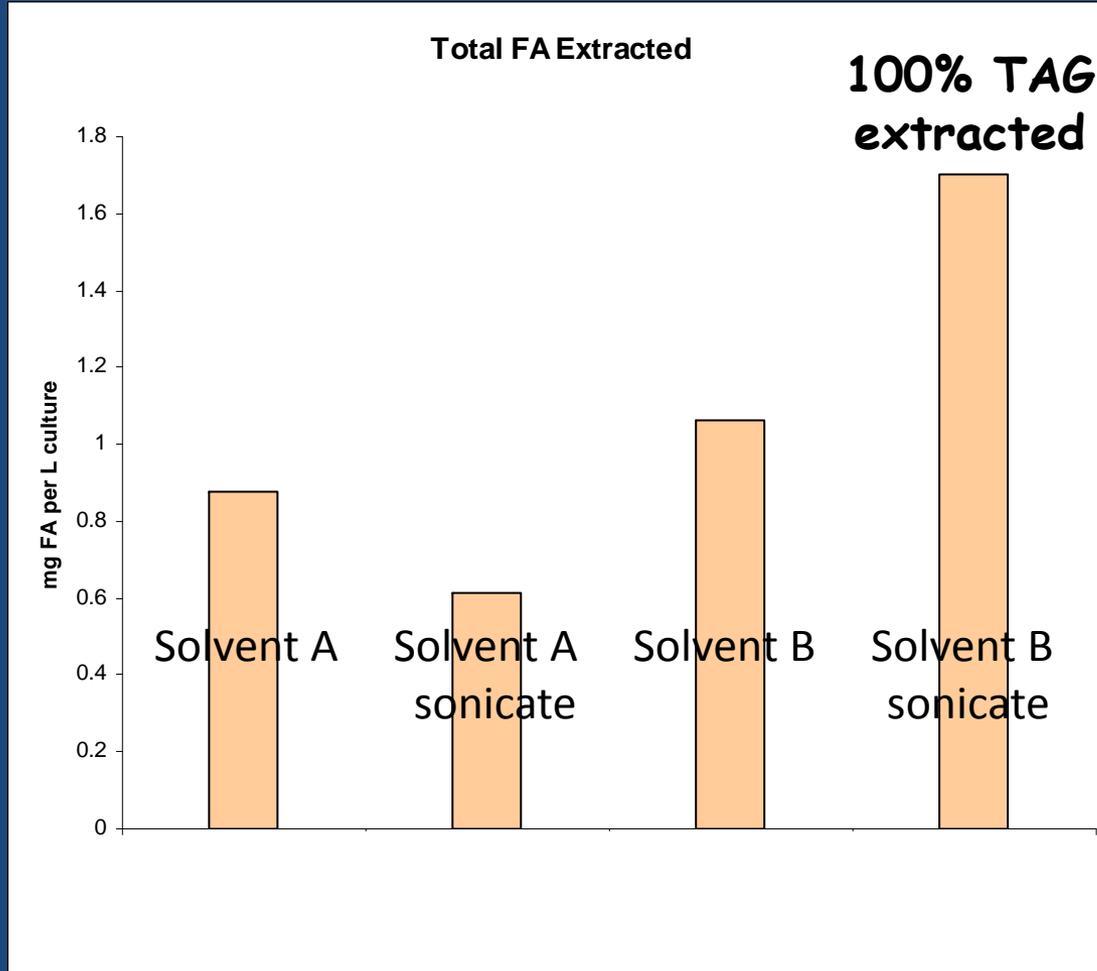
Cell survival after solvent extraction



Total cells/mL x 10^{-5} .

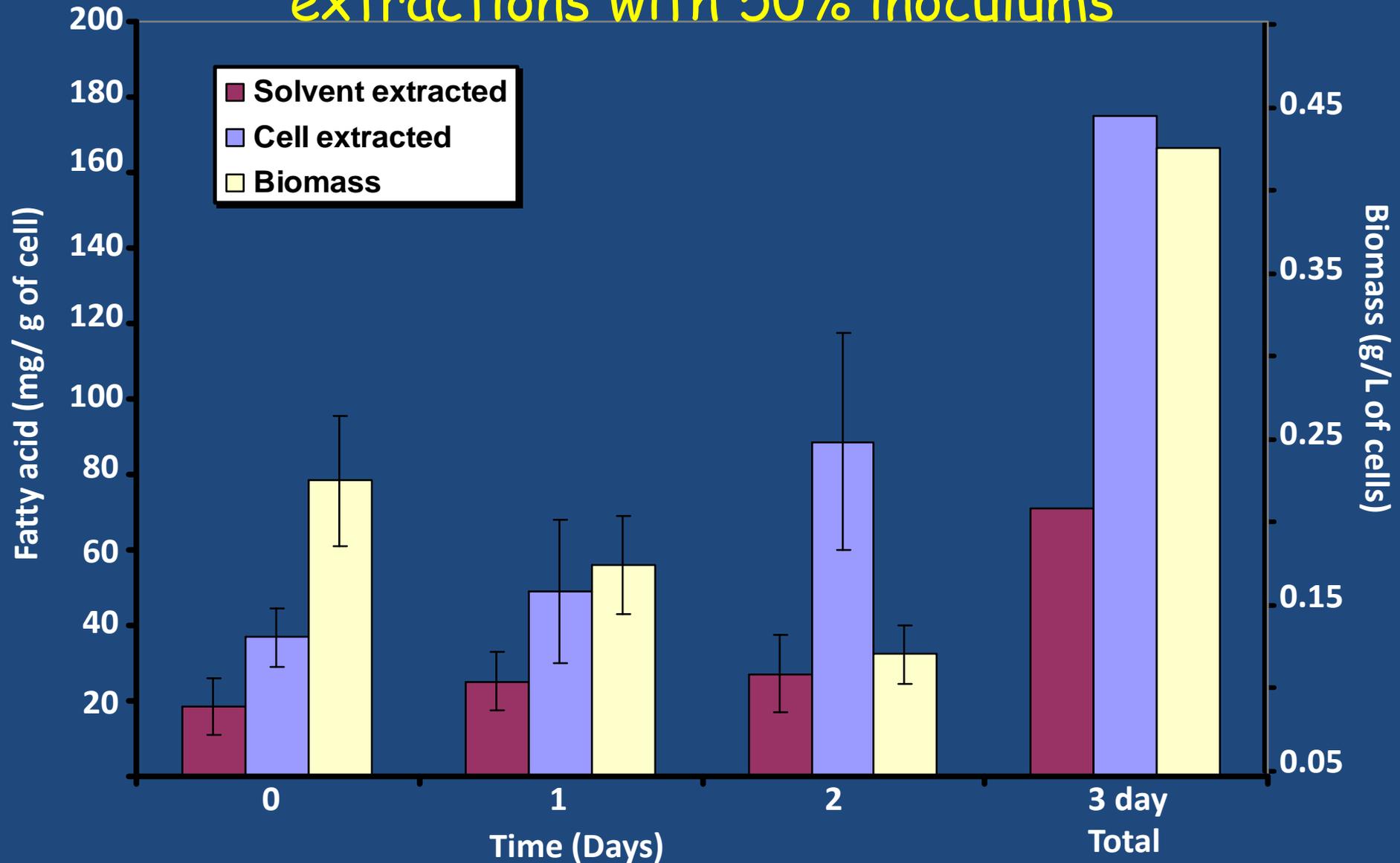
Incubation with bio-compatible organic solvents results in complete oil extraction from live cells

GC-MS
analysis



Ten minutes incubation, +/- 2 sec sonication

Optimizing solvent extraction; repetitive solvent extractions with 50% inoculums

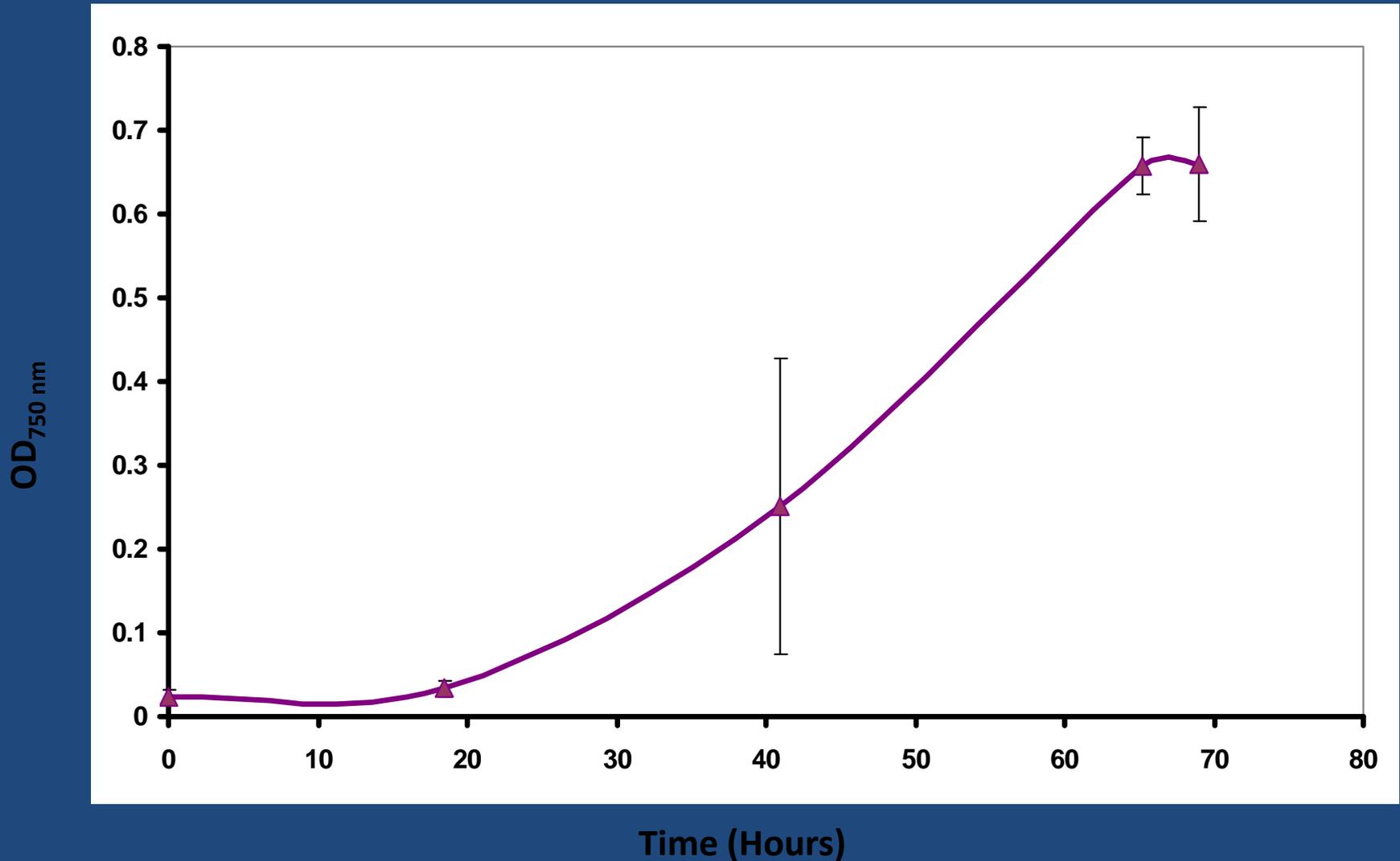


Repetitive solvent extraction yields more oil

Trait	Continuously extracted cultures, 3 day total	Batch extracted cultures; 3 day total
Biomass harvested	208 mg/L 2.4X	85 mg/L
Total lipids produced	175 mg/gdw 1.14 X	154 mg/gdw
Solvent extracted lipids	71 mg/gdw 1.39X	51 mg/gdw
% lipids solvent extracted	41%	33%

50% inoculum of continuously extracted cultures

Optimizing solvent extraction; repetitive solvent extractions with 10% inoculums



In contrast to a 50% inoculum, growth recovers with a 10% inoculum

Optimizing oil yield by solvent extraction, growth inhibition versus stress-induced oil production

- The use of large (50%) solvent-extracted inoculums reduces culture growth.
- Solvent extraction elevates algal oil production.
- The optimal inoculum concentration for solvent extraction systems remains to be determined.
- Solvent extraction may be used to harvest other non-polar molecules.
- Up to 40% of oils can be harvested without dewatering.
- Large inoculums can reduced contamination.

